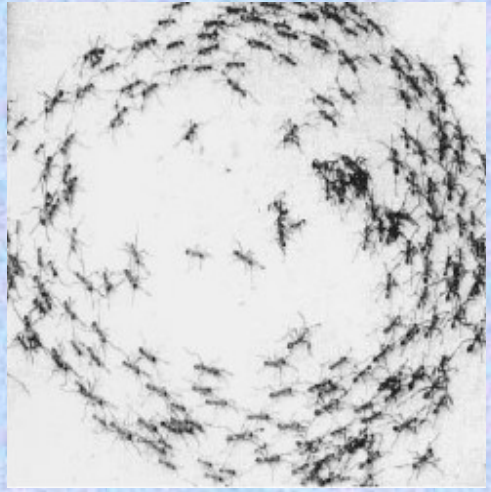
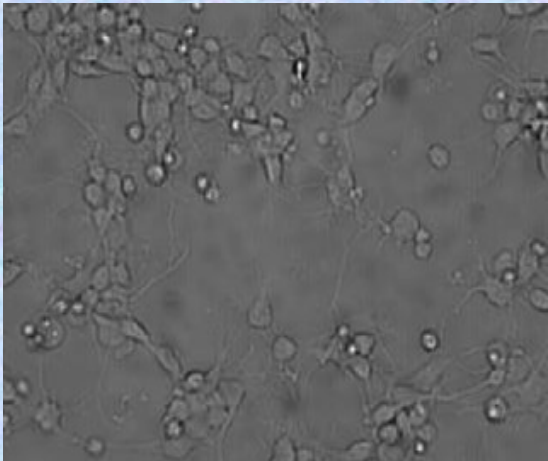
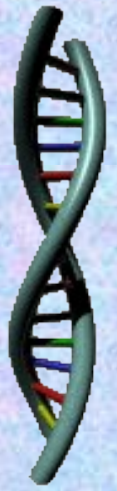


• What Physicists can do in Biology ?

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<http://www.phy.ncu.edu.tw/~ibp/>



Physics is vital in breakthrough in life sciences

- **Breakthrough in physical instrument:** optical microscope (Hooke, 1665), amplifier, X-ray, electron microscope, MRI, SPM, mass spectrometer, Single molecule microscopy,.....

Nobel laureates in physiology/medicine

that were physicists/had physics training:

- Georg von Békésy (physical mechanism of the cochlea, 1961)
- Francis Crick (DNA, 1962)
- Alan Hodgkin (nerve cell ,1963)
- Haldan Hartline (visual processes in the eye, 1967)
- Max Delbrück (bacteriophage ϕ ,1969)
- Rosalyn Yalow (radio-immunoassays of peptide hormones, 1977)
- Werner Arber (restriction enzymes , 1978)
- Erwin Neher (single ion channels in cells ,1991)
- Paul Greengard (signal transduction in nervous systems, 2000)
- Leland Hartwell (regulators of cell cycle, 2001)
- Peter Mansfield (NMR, 2003).....

Others: Schroedinger, Cooper,

Feigenbaum,...

What is Biophysics?

Biophysical Society defines as: "that branch of knowledge that applies the principles of **physics** and **chemistry** and the methods of **mathematical** analysis and **computer** modeling to understand how the mechanisms of biological systems work" .

• Why BioPhysics ?

- Material Nature of Bio-substances affect Biological properties. (Evolution made use of the physical properties of bio-materials)
- Physical principles & Laws holds from microscopic level → macroscopic level
- Traditional Biology is descriptive, non-quantitative

Why BioPhysics ?

- Physics is universal.
- Rise of molecular biology: DNA, RNA, protein, ATP... are **universal** in all living matters
- **Universality** in Central Dogma:
DNA → RNA → protein → Biological functions...
- New, interesting, exciting & useful.
- Lots of unsolved important problems.
- Techniques & Methodology in physics can probe the **fundamental principles** in bio-systems of a wide spectrum of scales in a **quantitative** way.

Era of modern Biophysics

- **Length Scales:**

nm □ μm ✕ mm □ cm ✕ m □ km

DNA, RNA, protein, intracellular, virus, bacteria, Intercellular, collective motion, insects, animals/plants, migration

- **Time Scales:**

fs ✕ ps □ ns ✕ ms □ s

e transfer, H-bonding, water DNA, RNA, protein rearrangement, protein folding DNA transcription

□ hr □ day □ year ✕ Byr

cell division Earth organisms, animal migration evolution

- **Knowledge: Interdisciplinary** 跨越各學科領域

Mathematics □ □ Physics □ □ Chemistry □ □ Biology □ □ Medical

BioPhysics ■ □ □ **Biology + Physics**

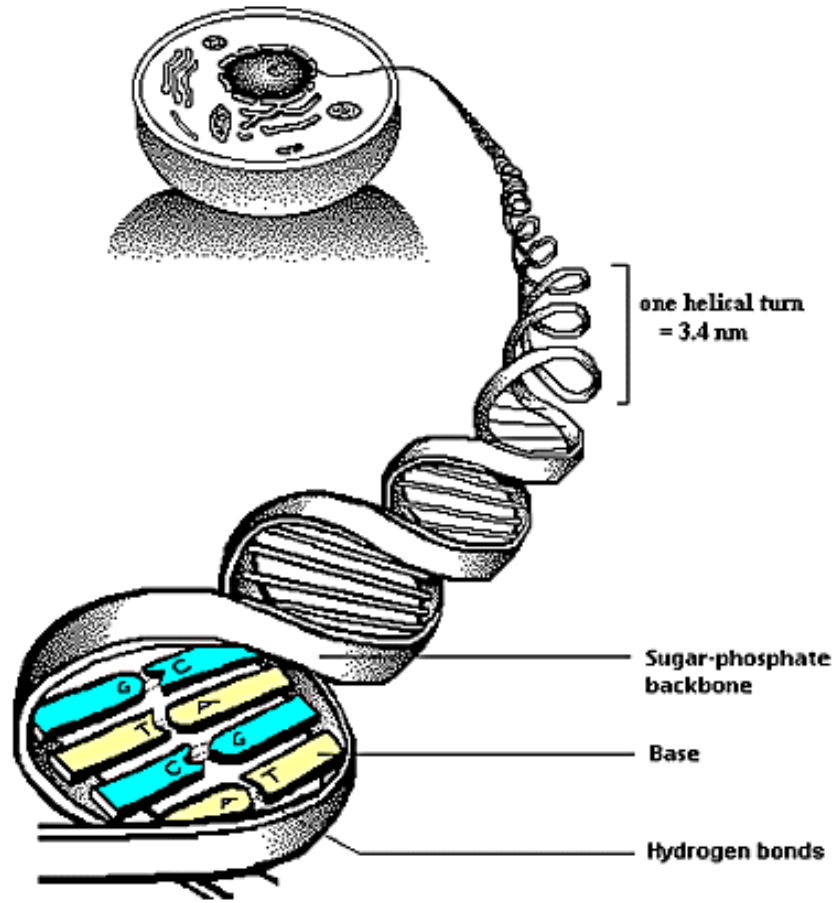
- Biophysicist is a TRUE Scientist ! Explore to the maximum freedom for doing science!

- 需要物理與非物理背景人材加入 !

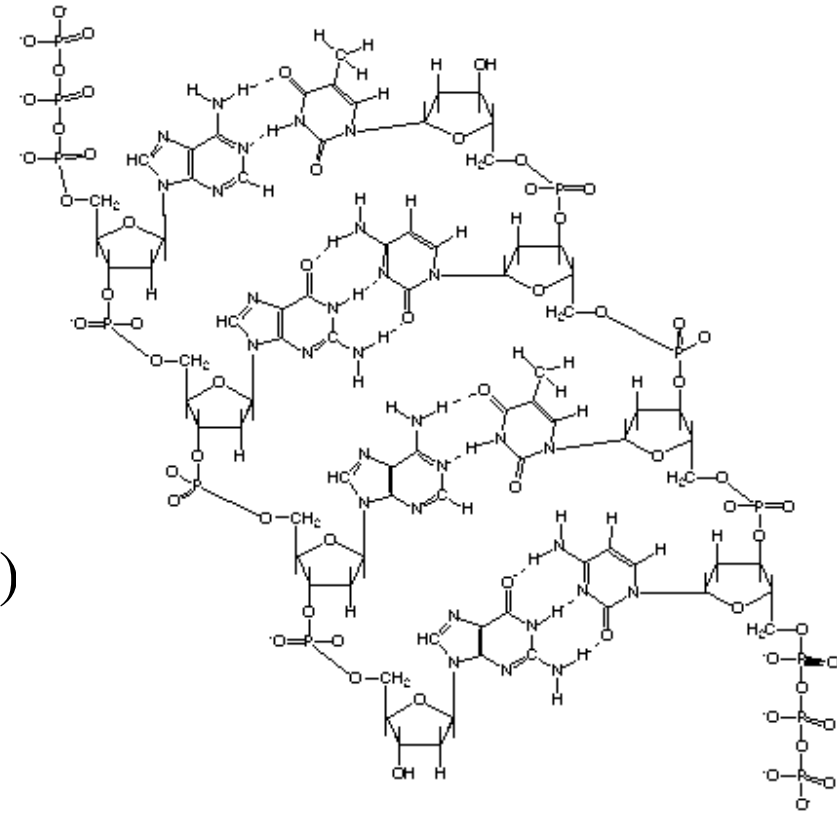
Elementary particles of Life

- Universal molecules: DNA, RNA, protein, ATP
- Interactions giving rise to bio-process: Central Dogma: DNA → RNA → protein → Biological functions...
- Nanomachines: molecular motors, FoF1 ATPase..
- How physical and chemical interactions lead to complex functions in cells ?
- Gene networks, protein networks

THE STRUCTURE OF DNA

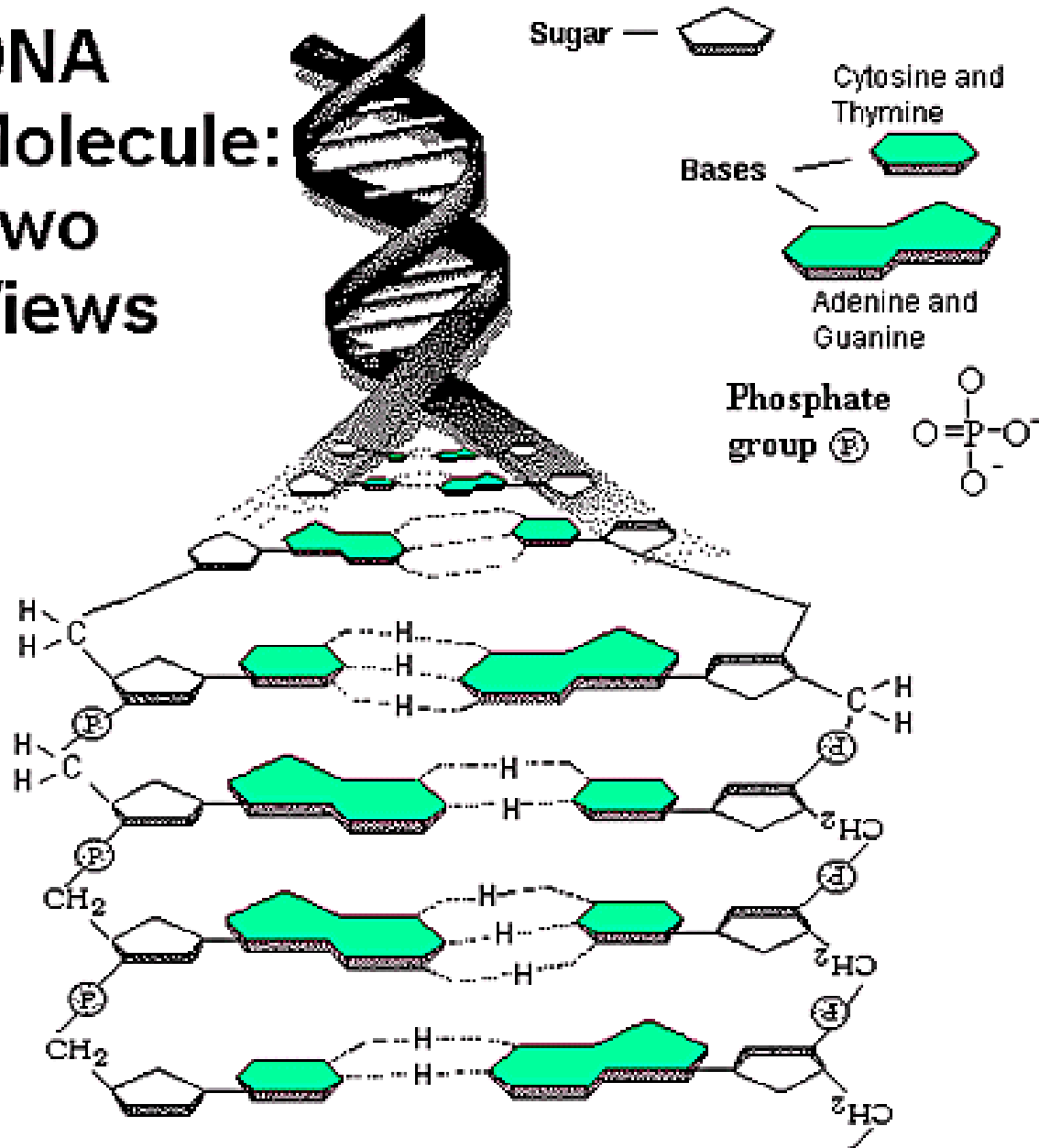


Cell
Nucleus
Chromosome
Chromatin



Double-stranded biopolymer, 2 sugar-phosphate chains (**backbones**) twisted around each other forming a RH (B-form) double helix.

DNA Molecule: Two Views



base pairs: **A-T** & **C-G**

Play (Torture) with DNA

- DNA stretching, elasticity
- DNA drag reduction
- DNA thermo-phoresis
- DNA condensation
- DNA under external fields
- DNA photolysis
- DNA ratchet motion
-

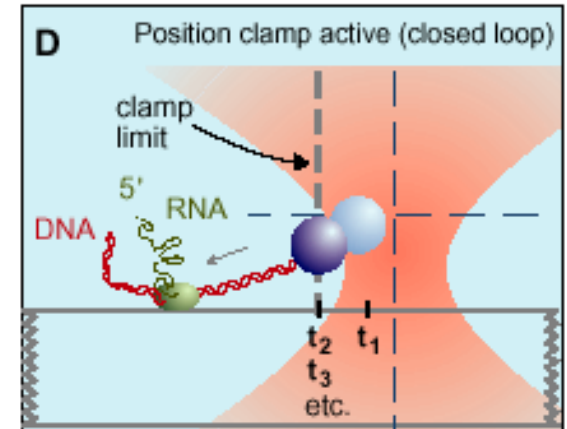
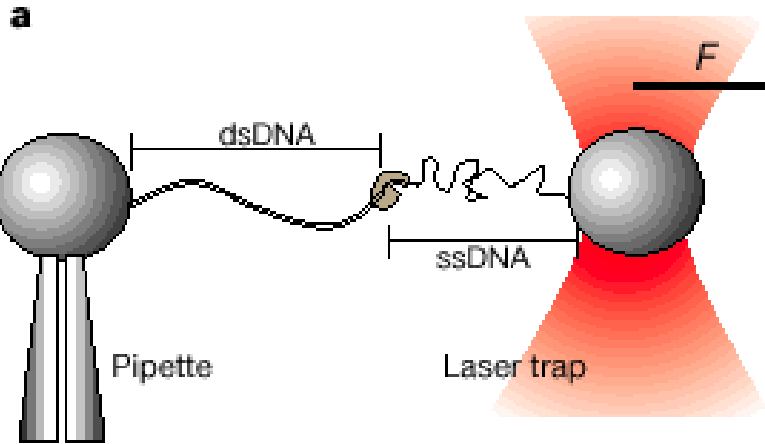


Mechanics/Elasticity of Single Bio-molecules

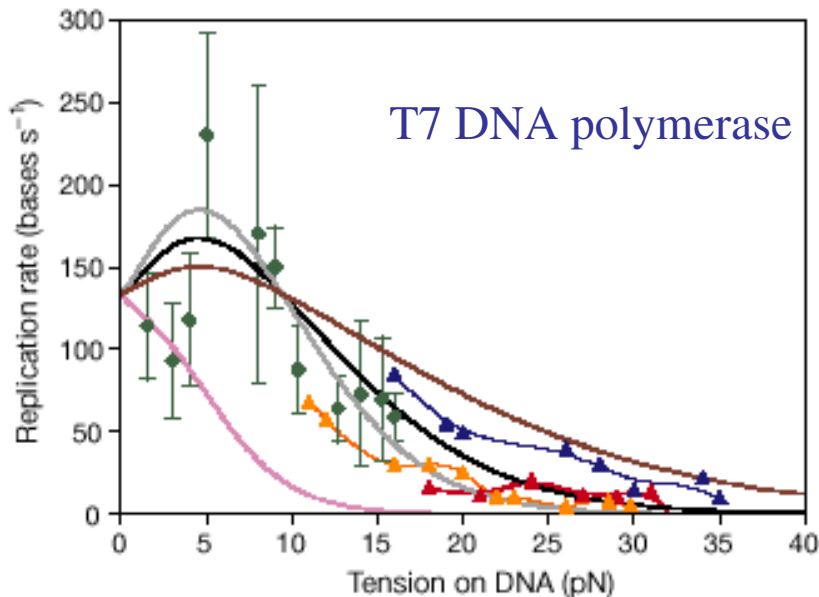
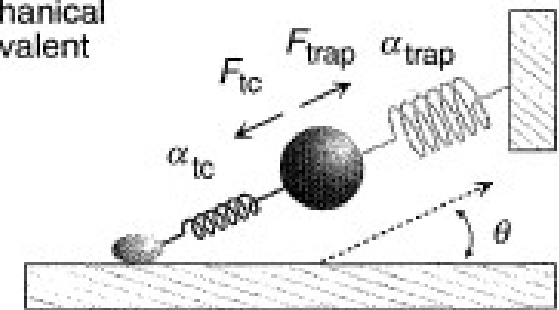
- To investigate the conformational changes in single bio-molecules, may provide significant insight into how the molecule functions.
- How forces at the molecular level of the order of pN underlie the varied chemistries and molecular biology of genetic materials?

DNA transcription by RNA polymerase

Bustemante et al, Nature 404, 103 (2000)

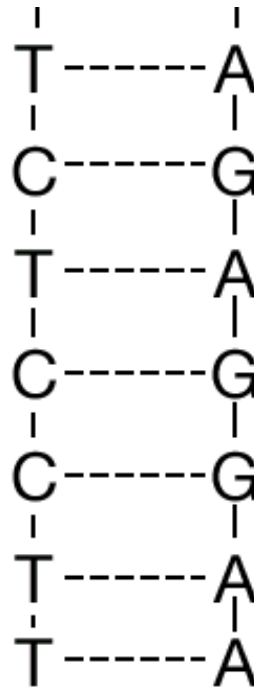
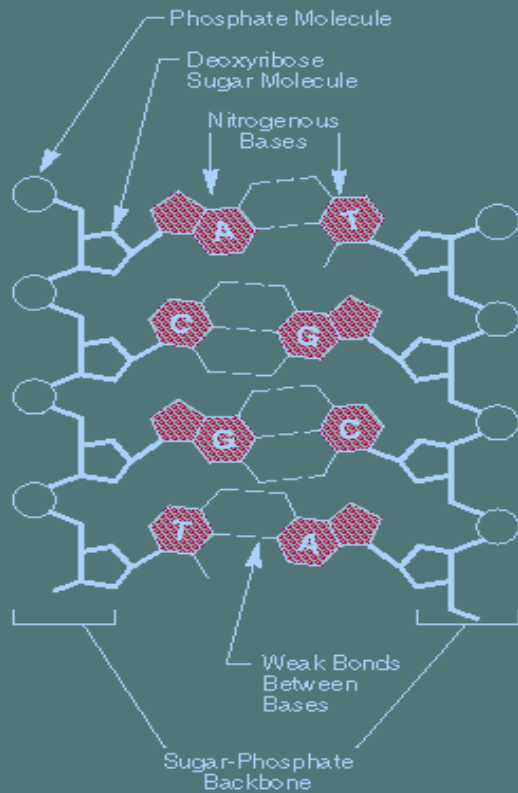


Mechanical equivalent



- effect of template tension polymerase activity
- Pausing & arrest during polymerase
- Mechanism of polymerization kinetics
- Tuning rate of DNA replication with external stresses

Physicist's view of the DNA chain



Double helix stabilized by H-bonds (bp interactions)

Polymer of persistence length $\sim 50\text{nm}$ under low force ($< 10\text{pN}$): Entropic elasticity. Complicated at high forces: cooperative behavior

Elasticity of dsDNA affect its structure and can influence the biological functions

Worm-like chain model (stiff chain)

$$\frac{E_{el}}{k_B T} = \frac{A}{2} \int_0^L ds \left(\frac{d\mathbf{t}}{ds} \right)^2 \quad \mathbf{t} = \frac{d\mathbf{r}(s)}{ds}$$

$|\mathbf{t}|=1$ inextensible

single strand

Rod-like chain model (twisted stiff chain)

Marko et al., Science 256, 506, 1599

(94);

$$\frac{E_{el}}{k_B T} = \frac{1}{2} \int_0^L ds \left[A \left(\frac{d\mathbf{t}}{ds} \right)^2 + C (\omega(s) - \omega_0)^2 \right] \quad \text{L 80, 1556 (98)}$$

Fitting from expts: $A=53\text{nm}$; $\frac{C}{A} = 1.64 \pm 0.04$ $\omega_0 = 2\pi / (10.5 \cdot 0.34) \text{ nm}^{-1}$

Can account for some supercoiling properties of DNA
Phenomenological model, no description of underlying
mechanism.

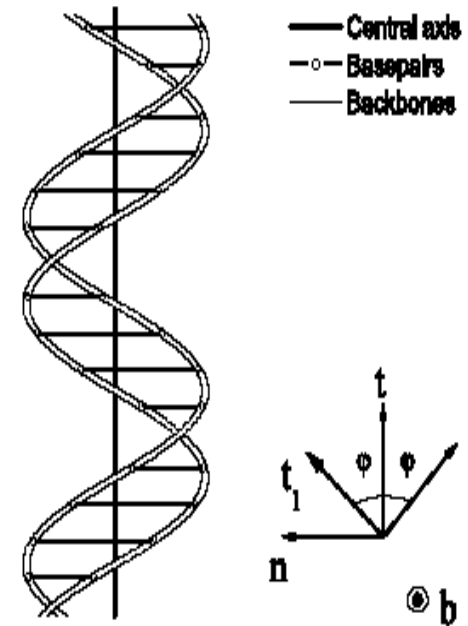
ZZO model for double-stranded DNA

H. Zhou, Z. Yang, Z.-c. Ou-Yang, PRL **82**, 4560 (99)

$$E_{\text{bs}} = \int_0^L \left[\frac{1}{2} \kappa \dot{t}_1^2 + \frac{1}{2} \kappa \dot{t}_2^2 + \rho(\varphi) \right] ds$$

$$= \int_0^L [\kappa \dot{t}^2 + \kappa \dot{\varphi}^2 + V(\varphi)] ds,$$

$$V(\varphi) = \frac{\kappa \sin^4 \varphi}{R^2} + \rho(\varphi),$$



$$\dot{\mathbf{r}} = \frac{1}{2}(\dot{\mathbf{t}}_1 + \dot{\mathbf{t}}_2) = \dot{t} \cos \varphi$$

φ =folding angle

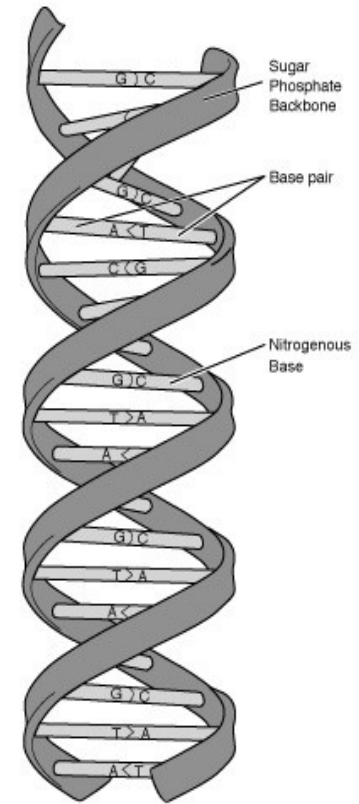
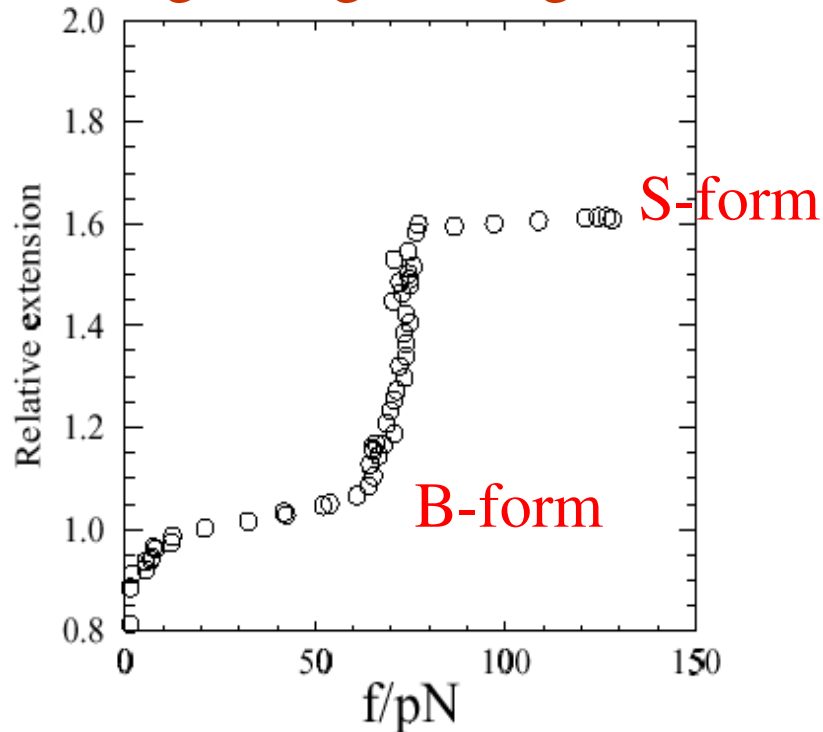
$$\rho(\varphi) = \begin{cases} \frac{\epsilon}{r_0} \left[\left(\frac{c_0}{\cos \varphi} \right)^{12} - 2 \left(\frac{c_0}{\cos \varphi} \right)^6 \right] & (\varphi > 0) \\ \frac{\epsilon}{r_0} (c_0^{12} - 2c_0^6) & (\varphi \leq 0) \end{cases}$$

B-form to S-form Transition under a Stretching force

Lai & Zhou, J. Chem. Physics 118, 11189 (2003)

Force Experiments

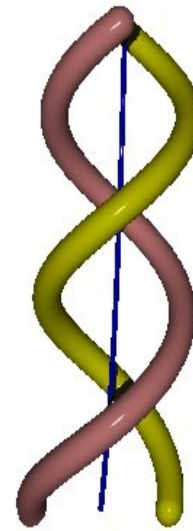
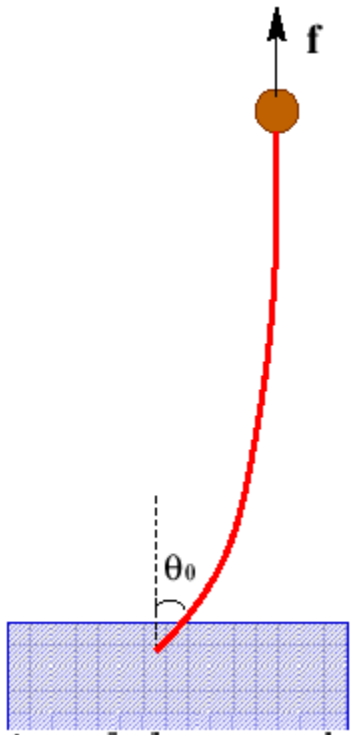
Stretching a single end-grafted DNA



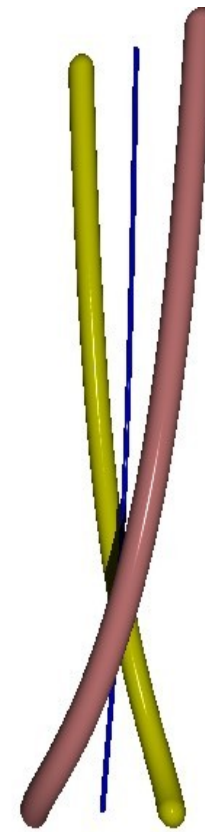
- Abrupt increase of 1.7 times in contour length of dsDNA near 65pN.
- Thermal fluctuations unimportant near onset of transition.

First order phase transition at β_t

First-order elongation: Stretch by untwisting



$\beta=0.073$



$\beta=0.075$

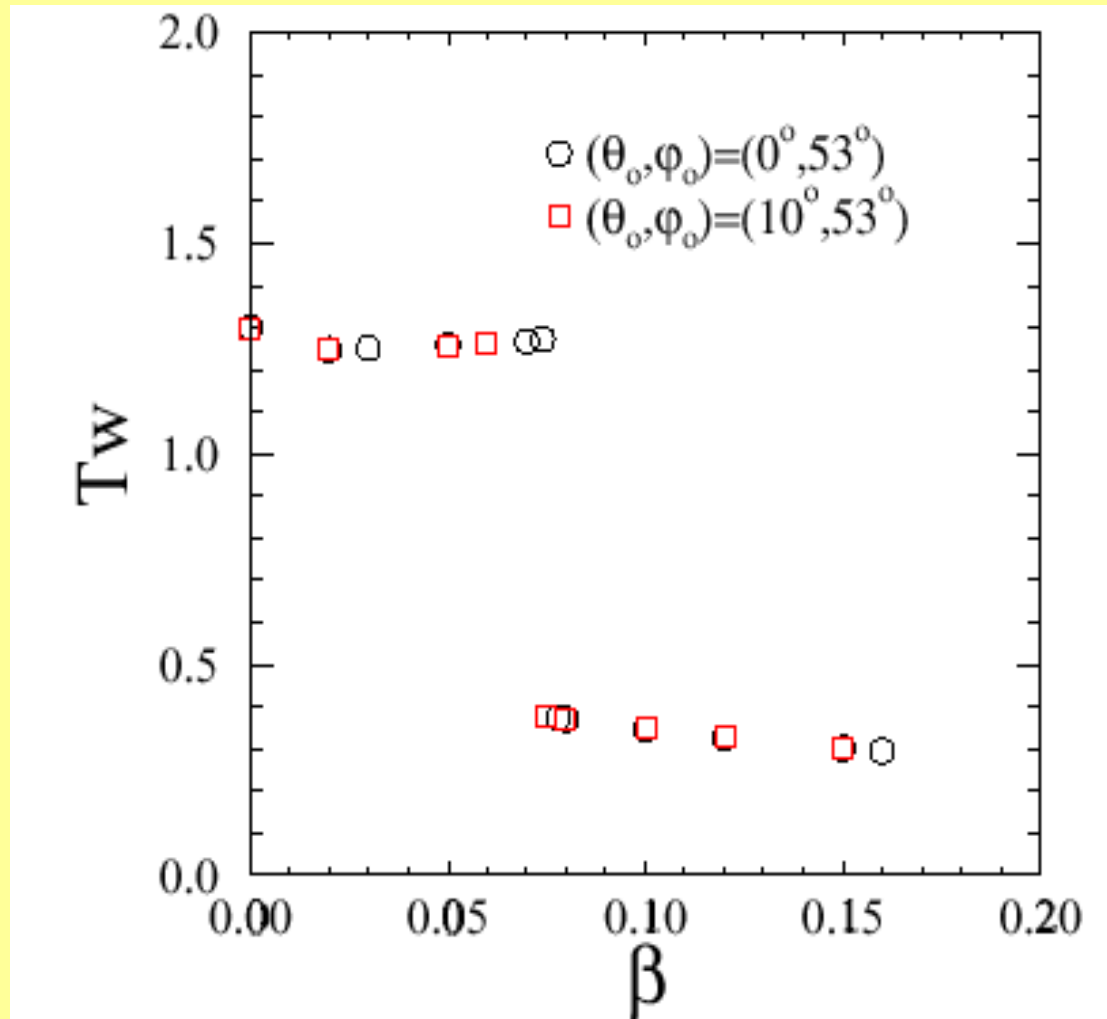
Untwisting upon stretching

Untwist per contour length from B-DNA, $\Delta Tw/L_0 \sim -100$ deg./nm;

- Almost completely unwound ~ 34 deg./bp
- Torque ~ 60 pN nm

Untwisting upon stretching

$$T_w = \frac{1}{2\pi} \int_0^L \sin \varphi ds$$

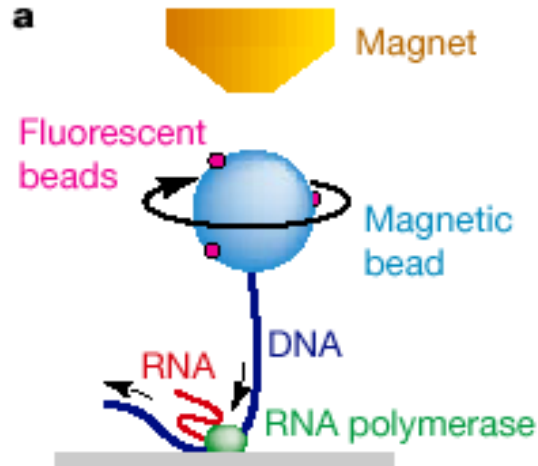


- Untwist per contour length from B-DNA, $\Delta T_w/L \sim -100$ deg./nm;
- Almost completely unwound ~ 34 deg./bp
- Torque ~ 60 pN nm

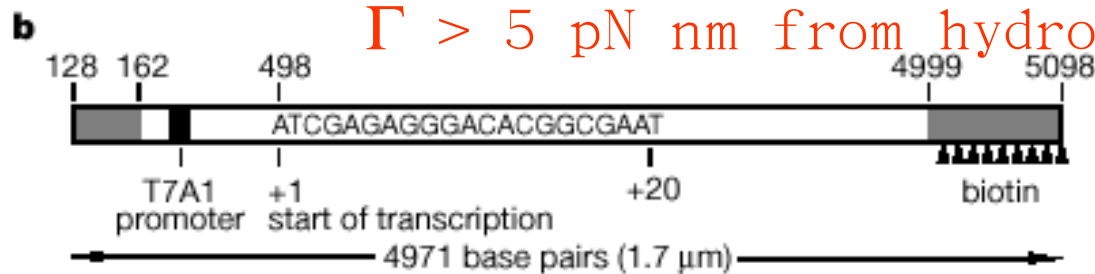
$$\bar{\Gamma} = \frac{\kappa}{R} \csc \varphi_t (V(\varphi_t) + 2\beta_t(1 - \cos \varphi_t))$$

Direct observation of DNA rotation during transcription by Escherichia coli RNA polymerase

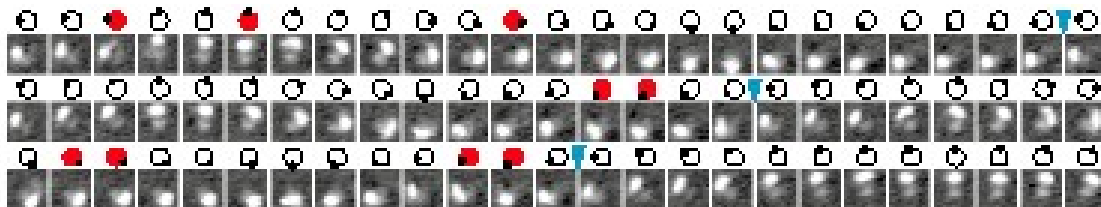
Harada et al., Nature 409, 113 (2001)



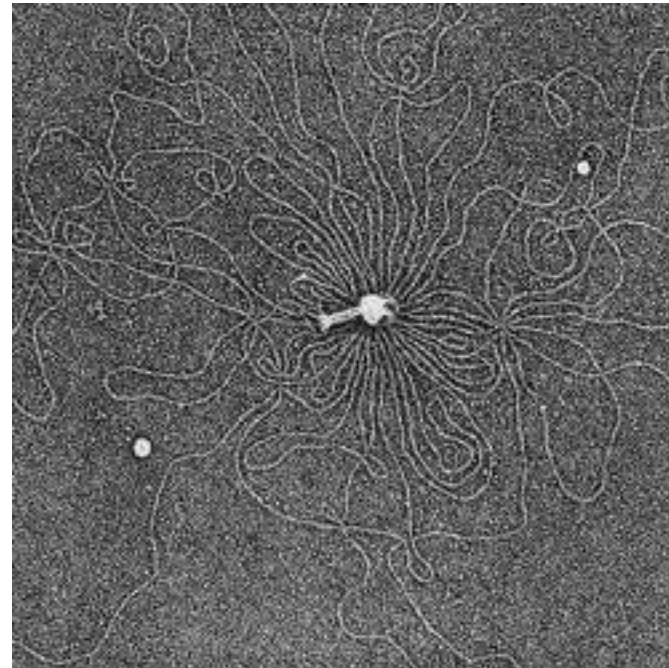
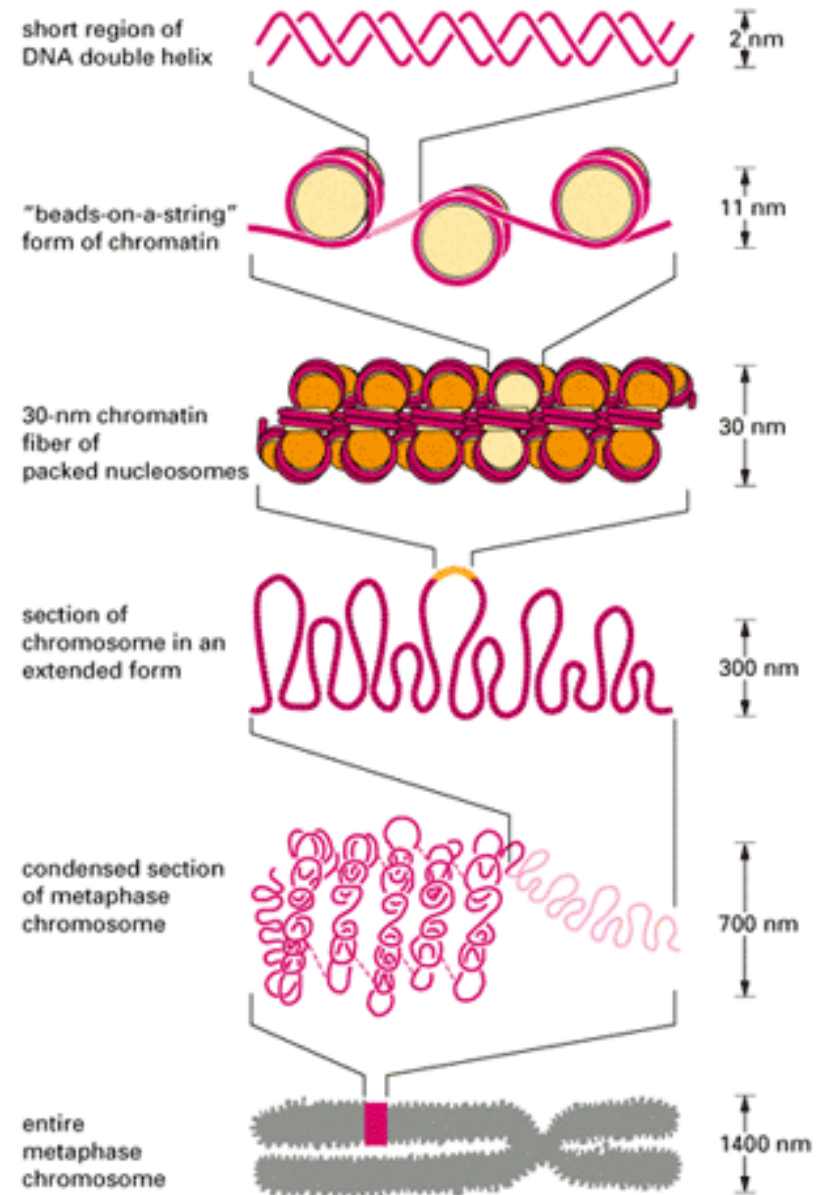
- DNA motor: untwisting gives rise to a torque
- B₀S transition provides a switch for such a motor.



$\Gamma > 5$ pN nm from hydrodynamic drag estimate

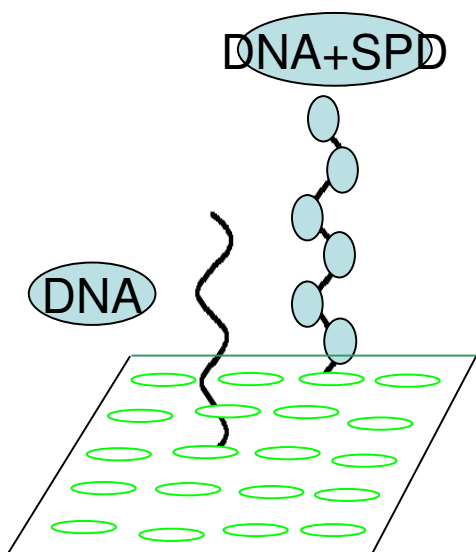
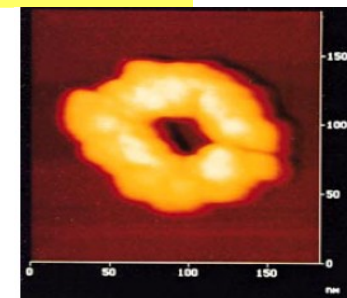
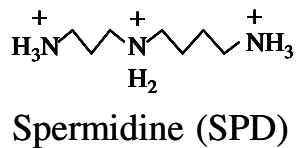
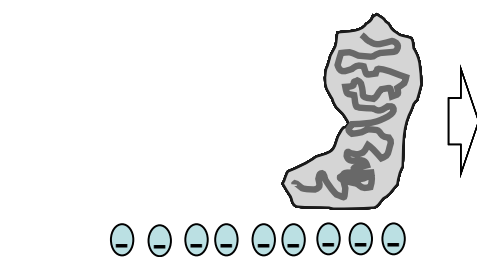


DNA condensation & packing



Complex competition of DNA elasticity, charge interactions, volume interactions, solvent effects.....

DNA condensed by spermidine



0.7% agarose gel



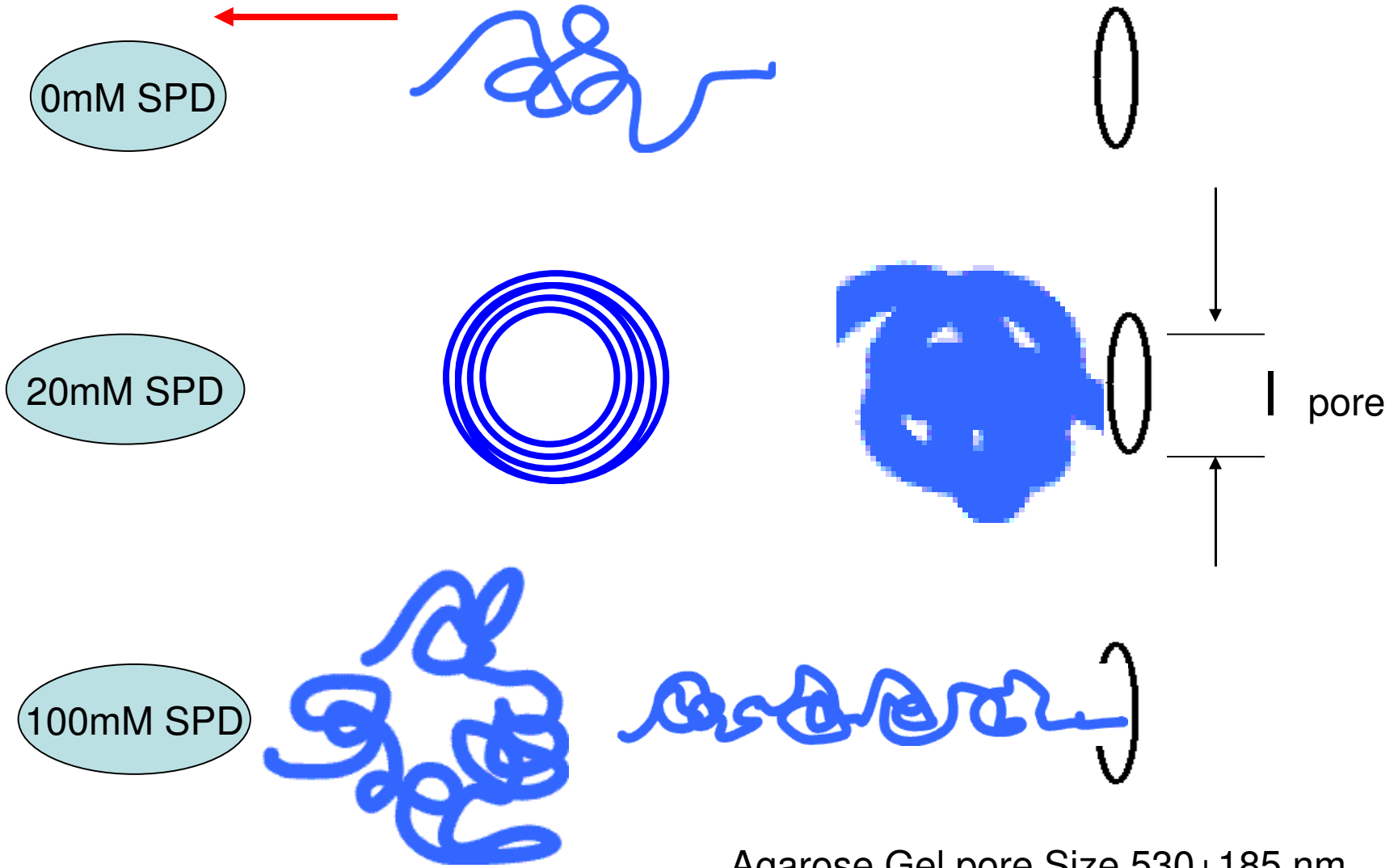
Single- λ DNA with
[SPD]=20mM jamming when
entering in 0.7% gel



Jamming: due to conformation changes in DNA

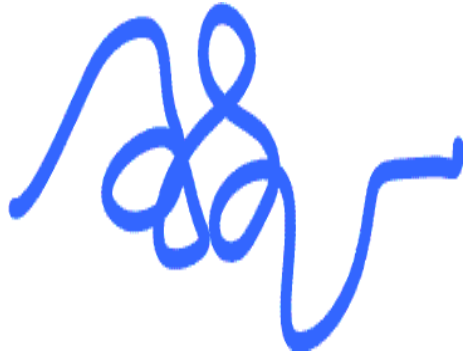
PRE 75, 041922 (2007)

E

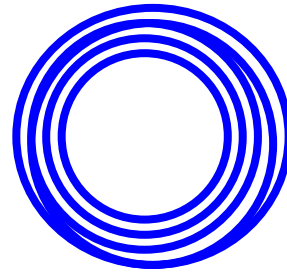


Agarose Gel pore Size 530 ± 185 nm
from AFM

De-Jamming: due to re-entrant condensation of DNA



Flexible coil (reptate thro' gel)



rigid condensed globule
(size > minimal pore size)



flexible again



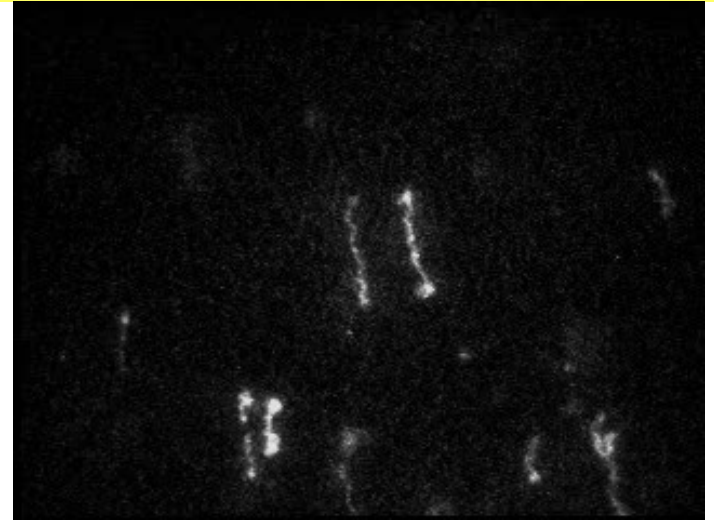
[SPD]

Re-entrant due to charge inversion (over-charge of DNA) ?

Controlled motion of DNA: external drives



In gel
under DC E field



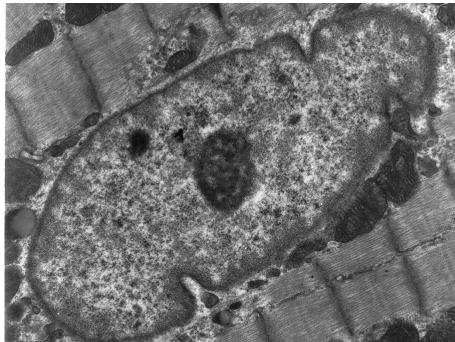
DNA ratchet motion under AC electric field



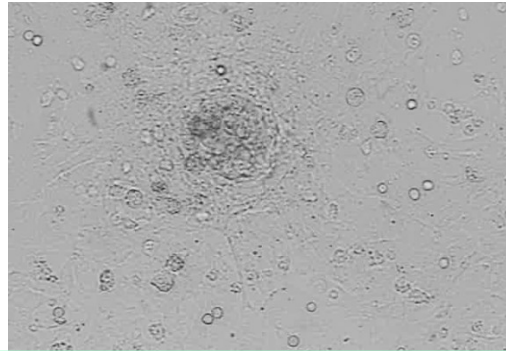
Simple to Complex: emerging properties in bio-systems

Couplings, interactions, nonlinearity, feedback... → collective behavior, bio-functions

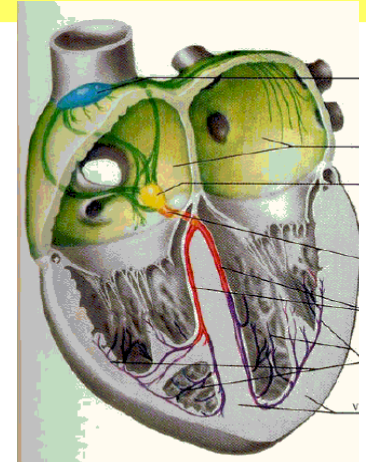
(I) cardiac cells → Heart



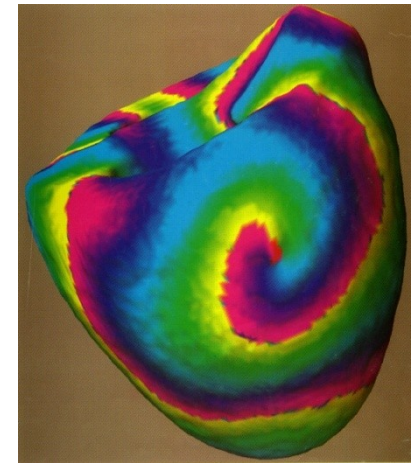
Cardiac myocyte



Synchronized beating of myocytes



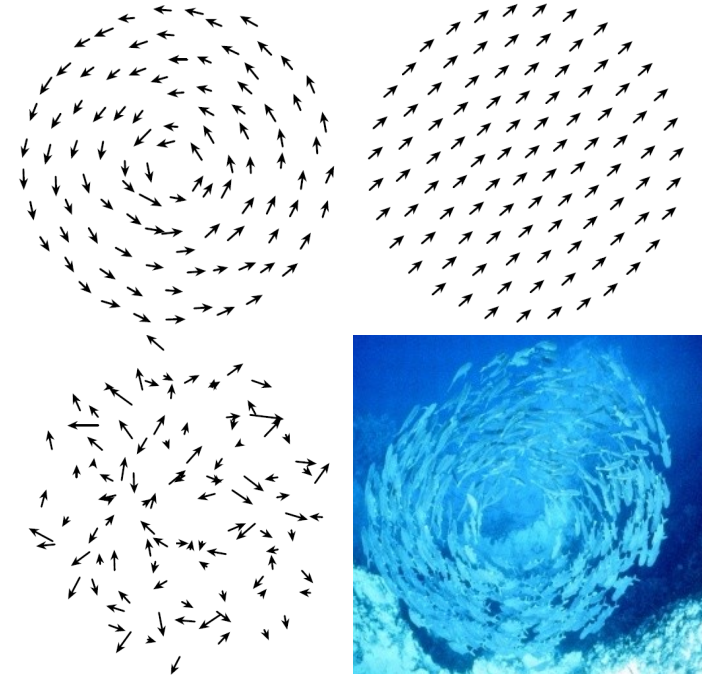
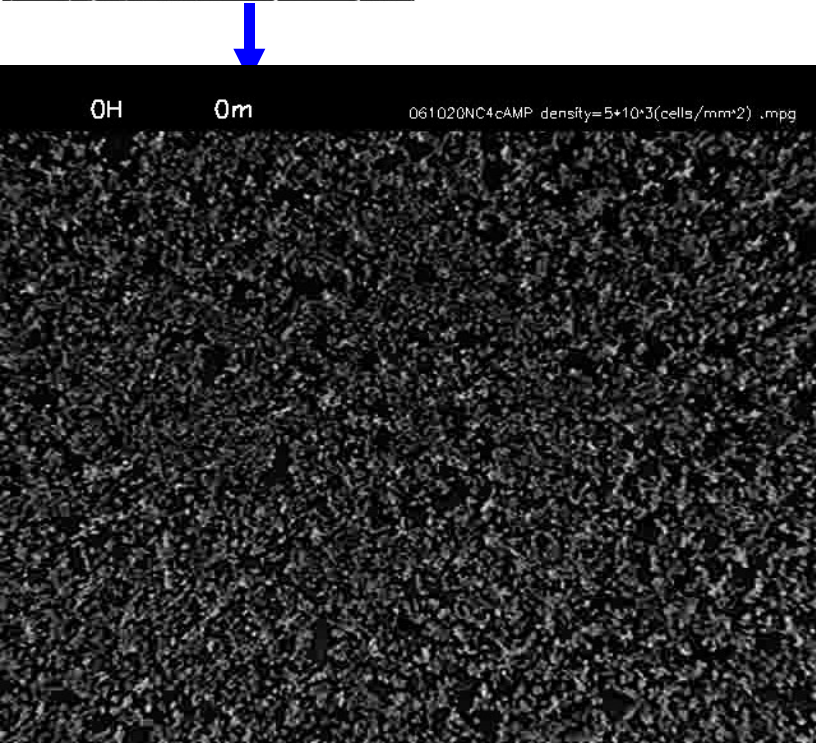
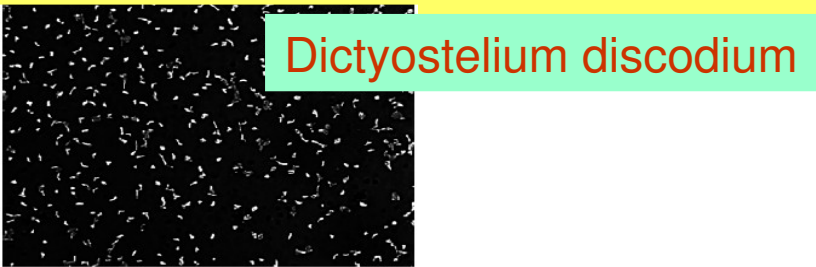
spiral waves:



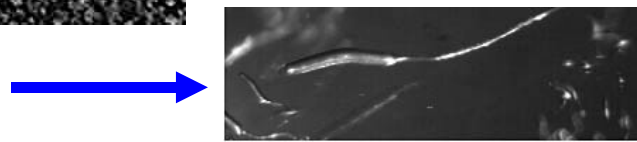
Coupled oscillator networks of Cardiac cells:
nonlinear dynamics, spiral waves, spatio-temporal patterns...

Simple to Complex: emerging properties in bio-systems

(II) Single cell/organism → collective motion

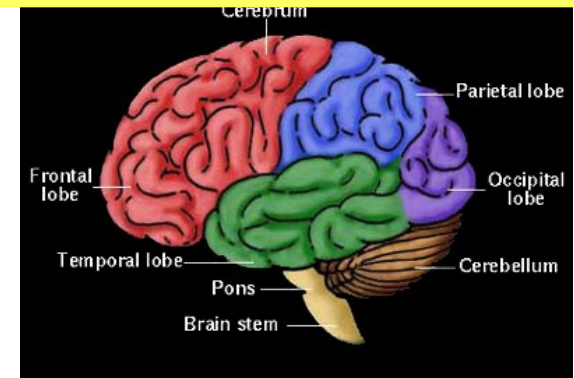
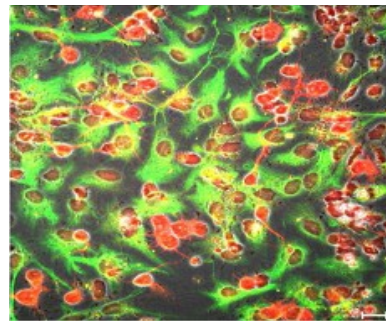
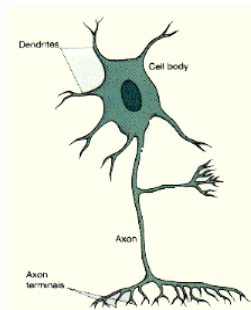


物種之群體運動理論：
魚群、昆蟲、細菌之習
體運動模式



emerging properties in bio-systems

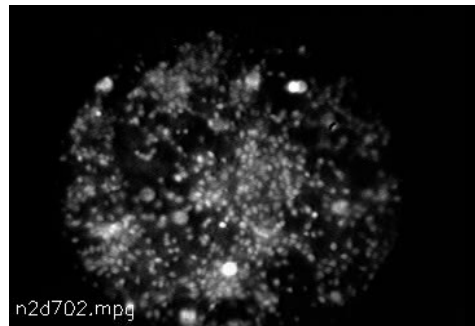
(III) Neurons \square Network \square Brain \square Behavior



Hodgkin-Huxley Model
(1952)

Network connection: **synapses**

Neuro/cognitive science



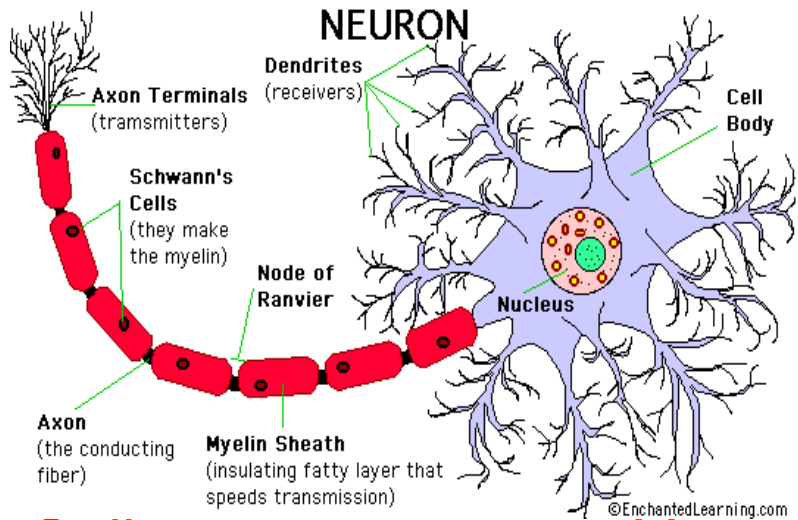
Synchronized Firing

Complex behavior/function determined by neuron connections.

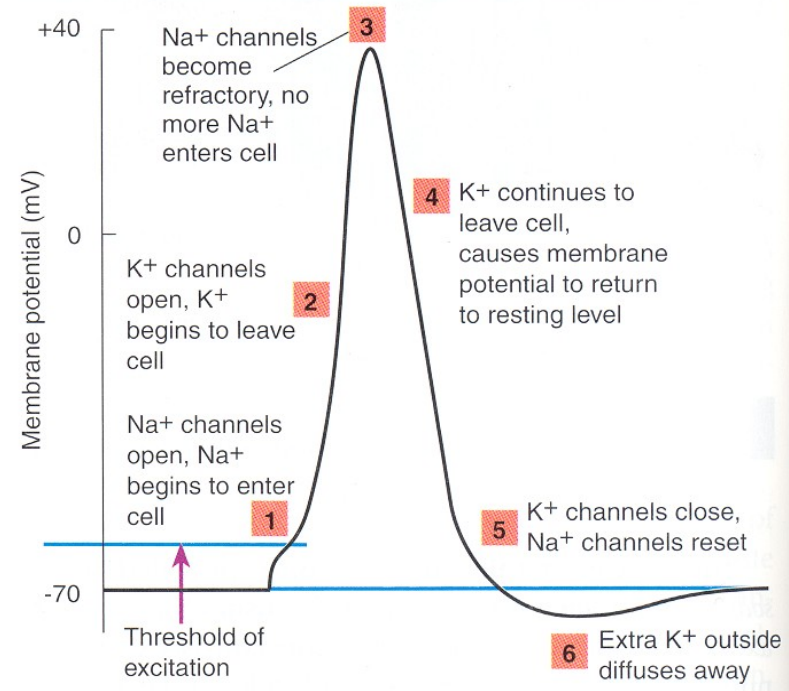
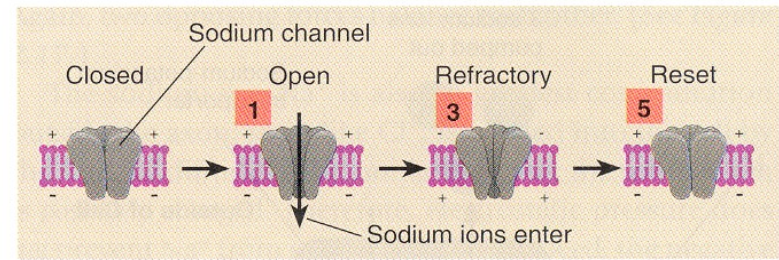
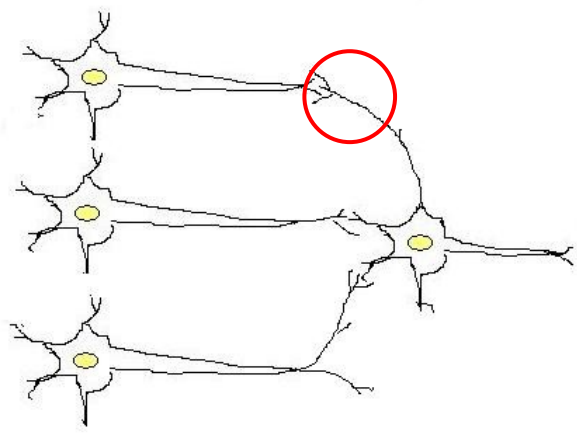
Complex neuronal Network:

- A single neuron in vertebrate cortex connects ~ 10000 neurons
- Mammalian brain contains $> 10^{11}$ interconnected neurons
- Signal & information convey via neuronal connections—coding

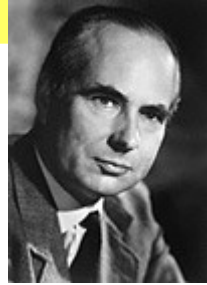
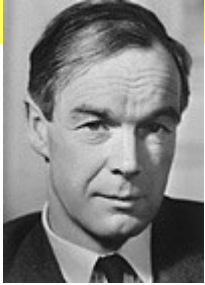
Neuron & Action Potential



Spike: ~ 1 ms, 100mV
Propagates along the axon to the junction of another neuron
---synapse

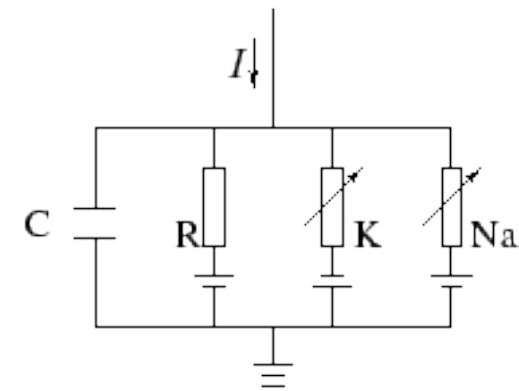
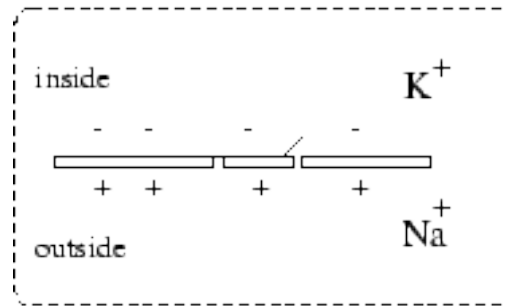


Hodgkin-Huxley model (1952)



Expts. On giant axon of squid:
time & voltage dependent Na, K ion channels
+ leakage current

$$I(t) = I_C(t) + \sum_k I_k(t)$$



$$\sum_k I_k = g_{Na} m^3 h (u - E_{Na}) + g_K n^4 (u - E_K) + g_L (u - E_L).$$

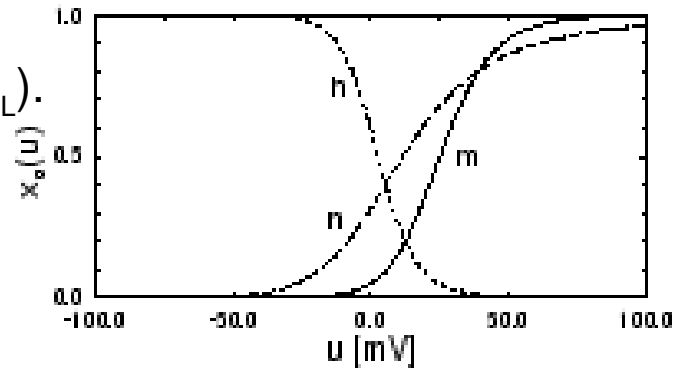
gating variables:

$$\dot{m} = \alpha_m(u) (1 - m) - \beta_m m$$

$$\dot{n} = \alpha_n(u) (1 - n) - \beta_n n$$

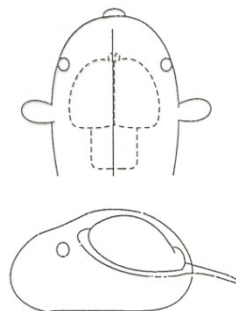
$$\dot{h} = \alpha_h(u) (1 - h) - \beta_h h$$

empirical functions

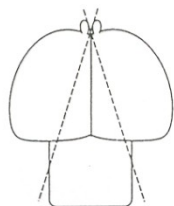


Experiments

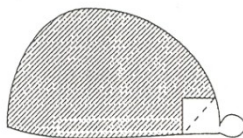
Schematic procedures in preparing the sample of neuron cells from cerebral cortex embryonic rats



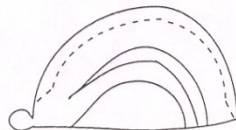
1. 剪開幼鼠頭部, 取出大腦



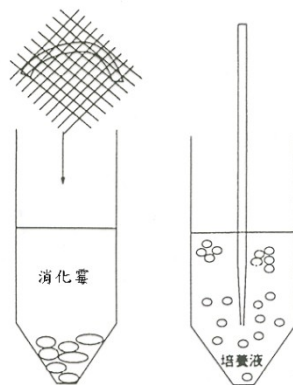
2. 剪下左右半腦



3. 剝去腦之保護膜

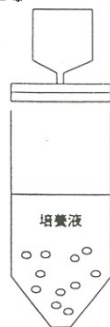


4. 剪下大腦皮層部份

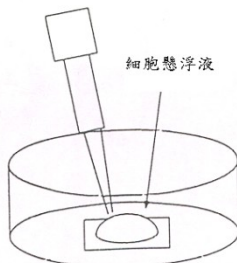


5. 加入消化酶

6. 加入培養液



7. 過濾雜質



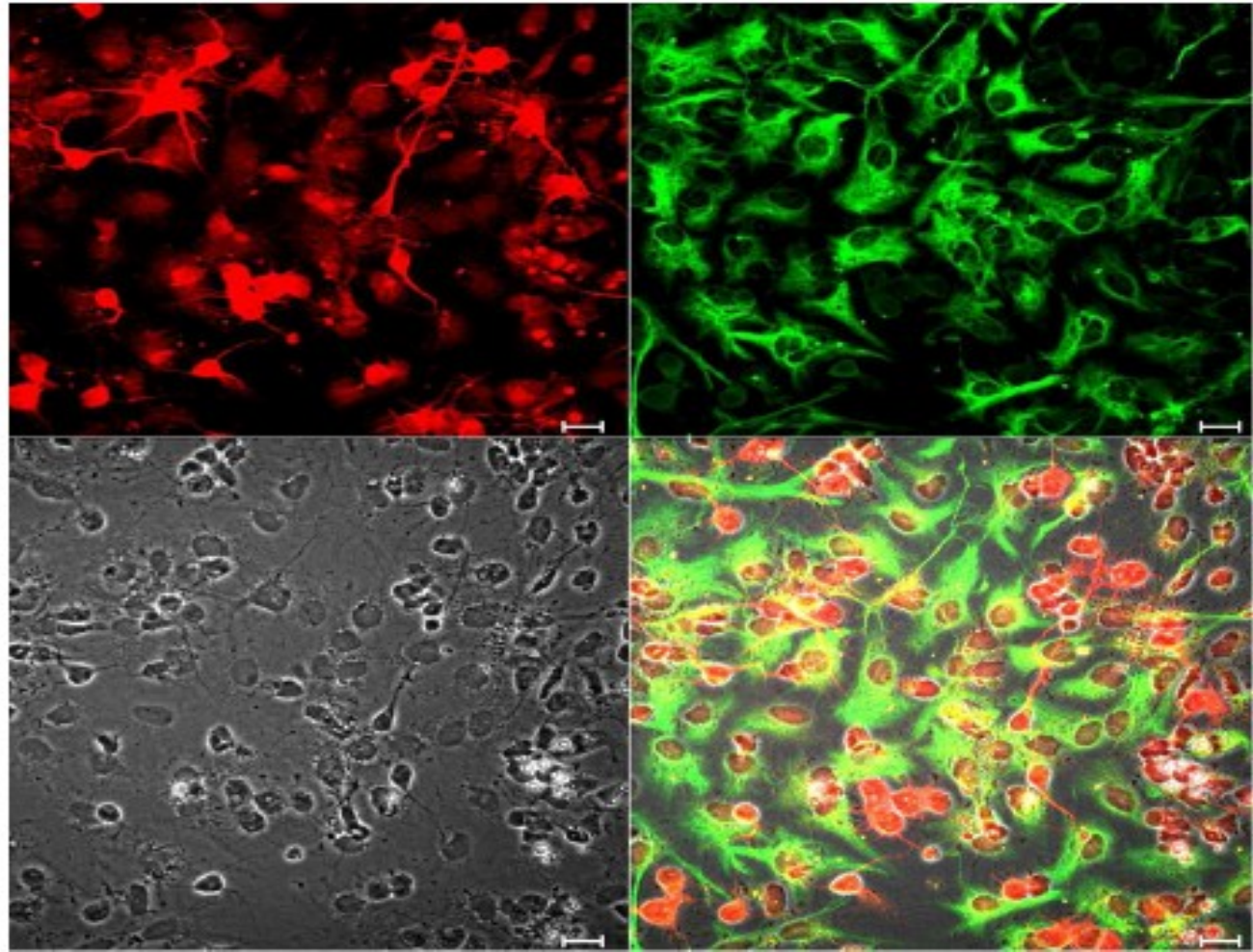
8. 將細胞懸浮液打入培養皿

Embryos of Wistar rats
E17~E18 breeding days



<http://mouse.kribb.re.kr/mousehtml/kistwistar.htm>

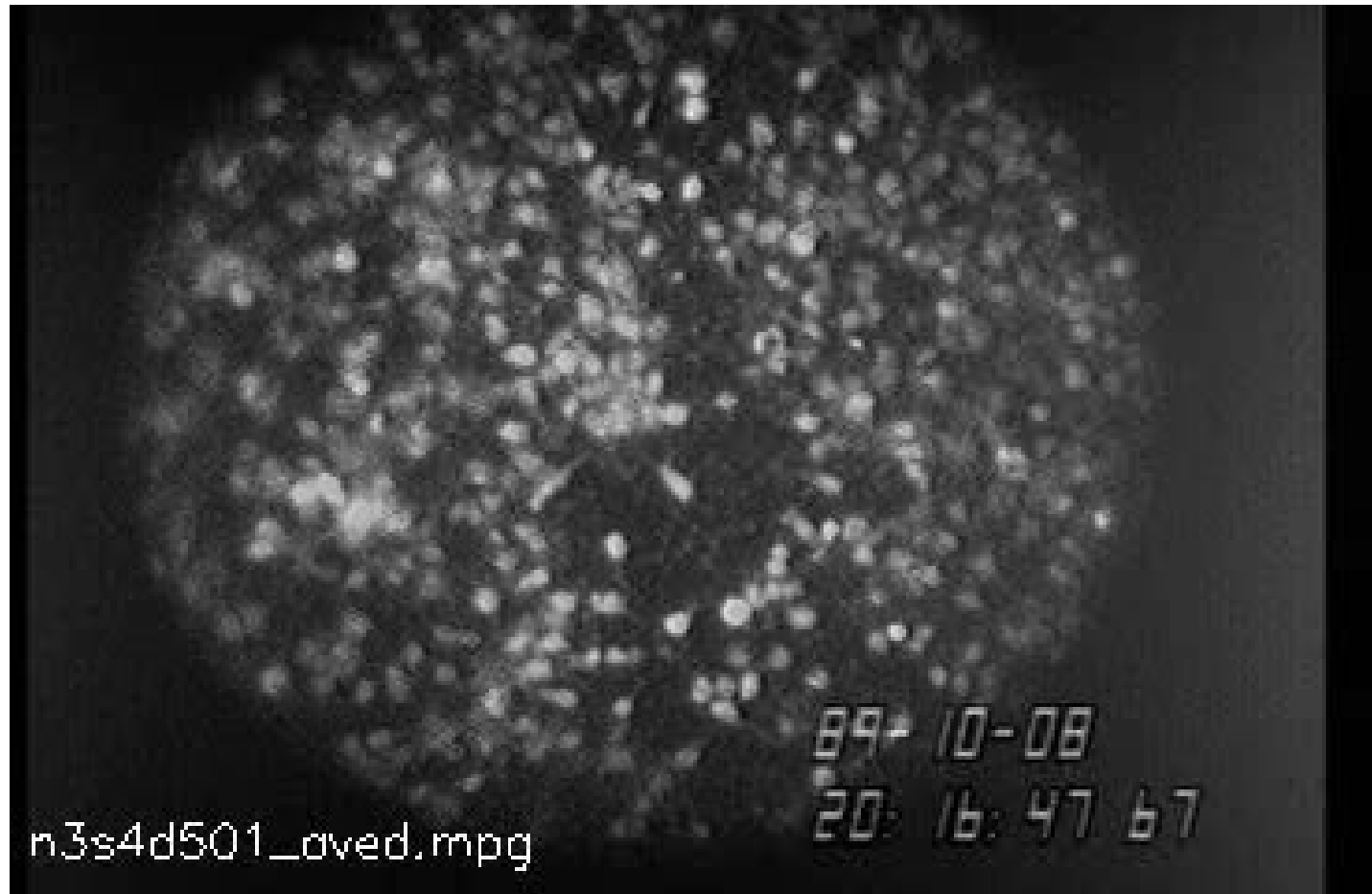
Growth of axon connection to form a network



Typical confocal microscope pictures of cultures used in our experiments.
Red: anti-MAP2 (neuronal marker); Green, anti-GFAP (glia marker). Black & white:
phase contrast image; Merge of the three images above.

Optical recording of fluorescence signals from firing network

Firing of the network is monitored by the changes in intracellular $[Ca^{2+}]$ which is indicated by the fluorescence probe (Oregon Green).

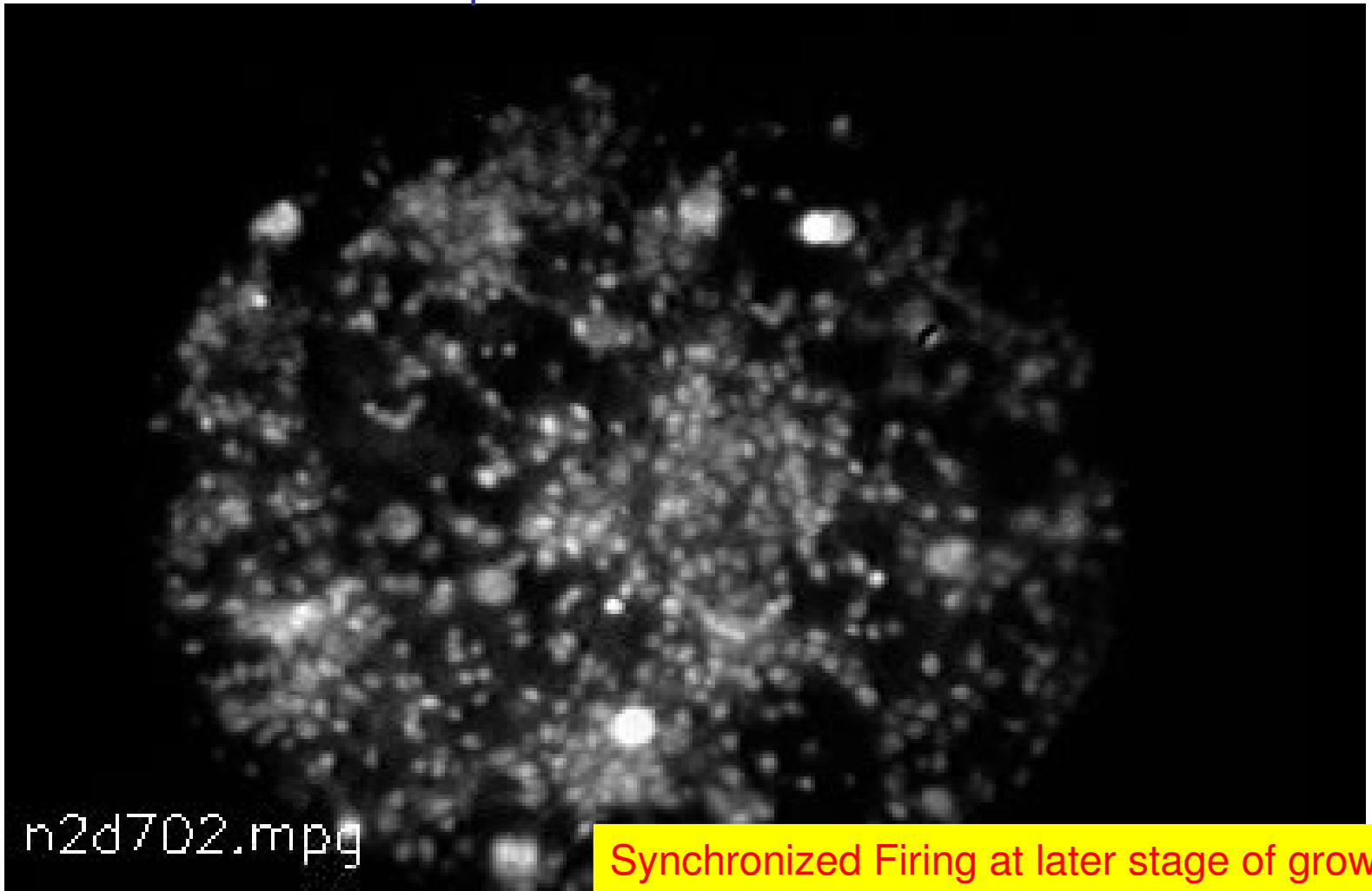


Non-synchronous Firing in early stage of growth

Synchronized Firing of Neuronal Network Culture

Spontaneous firing of the cultures are induced by reducing $[Mg^{2+}]$ in the Buffered salt solution

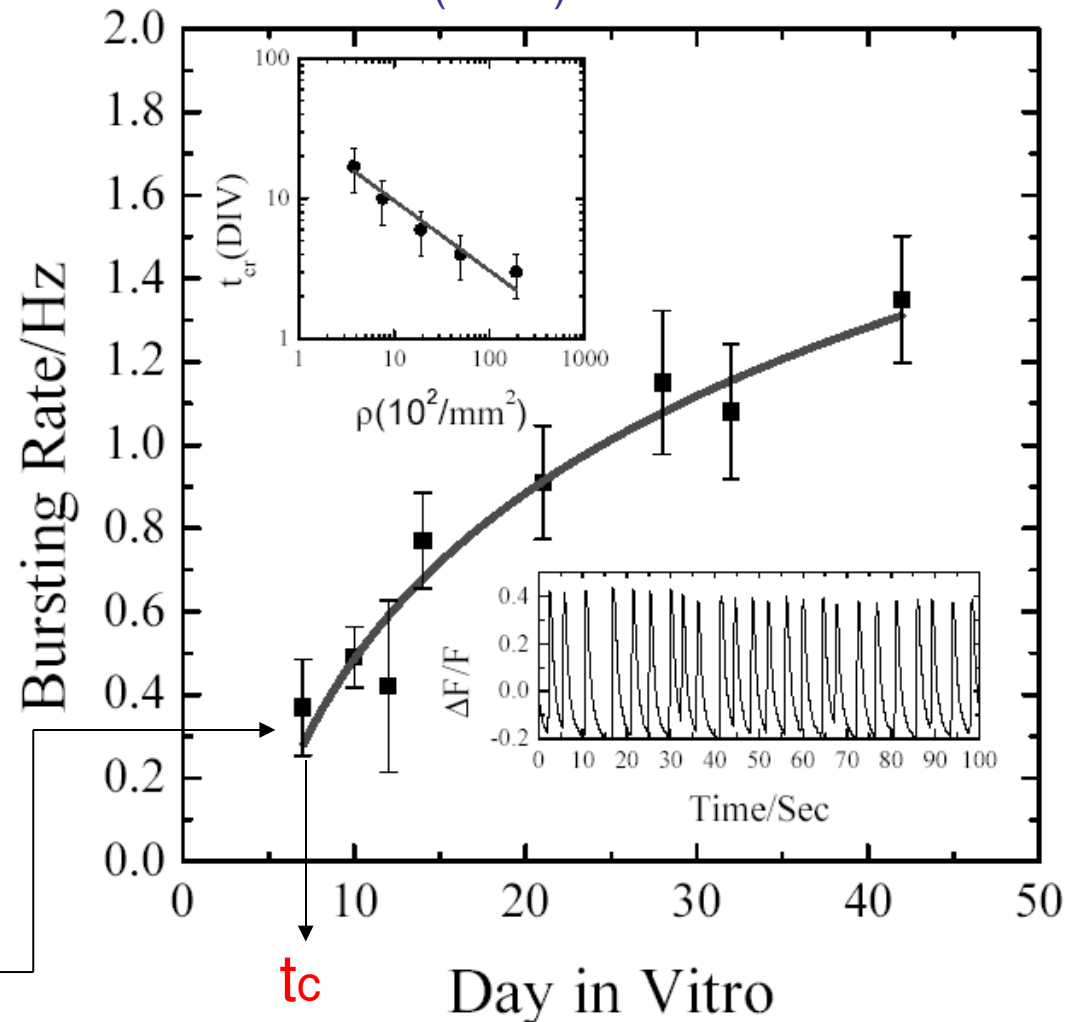
Firing \rightarrow the changes in intracellular $[Ca^{2+}]$ indicated by the fluorescence probe.



Time dependence of the SF frequency for a growing network

Phys. Rev. Lett. 93 088101 (2004)
PRE (2006)

- Critical age for SF, t_c
- SF freq. grows with time
 $f = f_c + f_0 \log(t/t_c)$

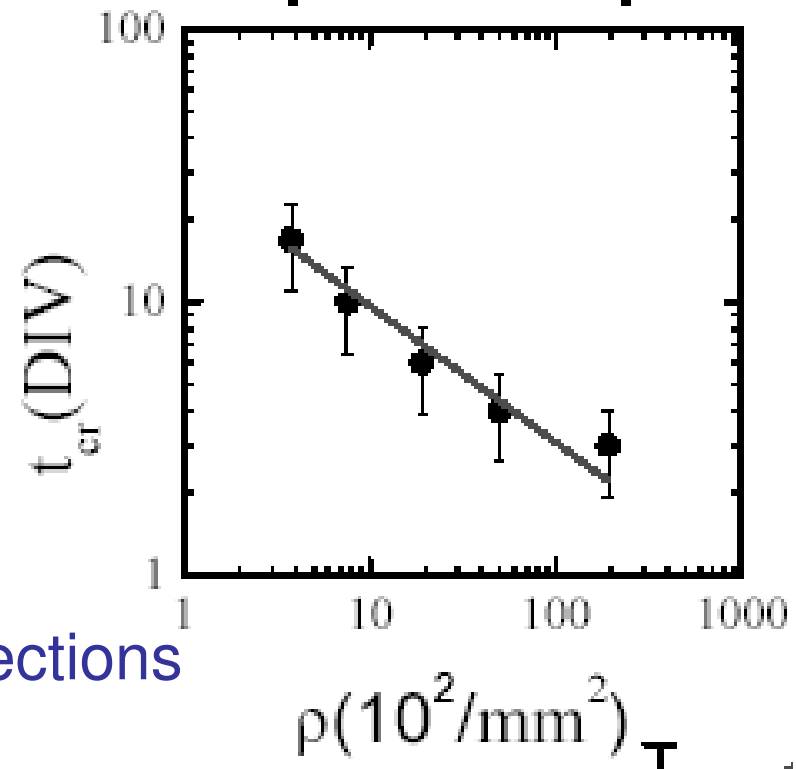


Onset time for SF as a function of cell density

- Critical age for SF

$$t_c \propto \frac{1}{\sqrt{\rho}}$$

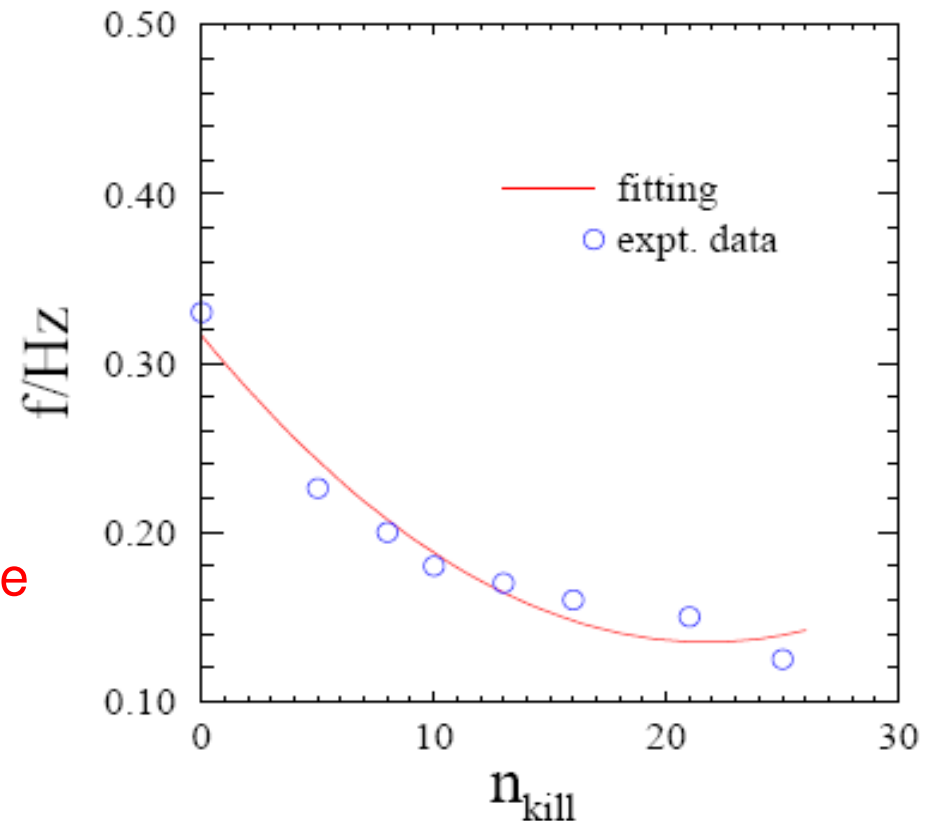
- $f = f_c + f_0 \log(t/t_c)$
- f increases with the effective connections
- f_c is indep. of ρ



Synchronous firing frequency $f \sim$ mean connectivity k

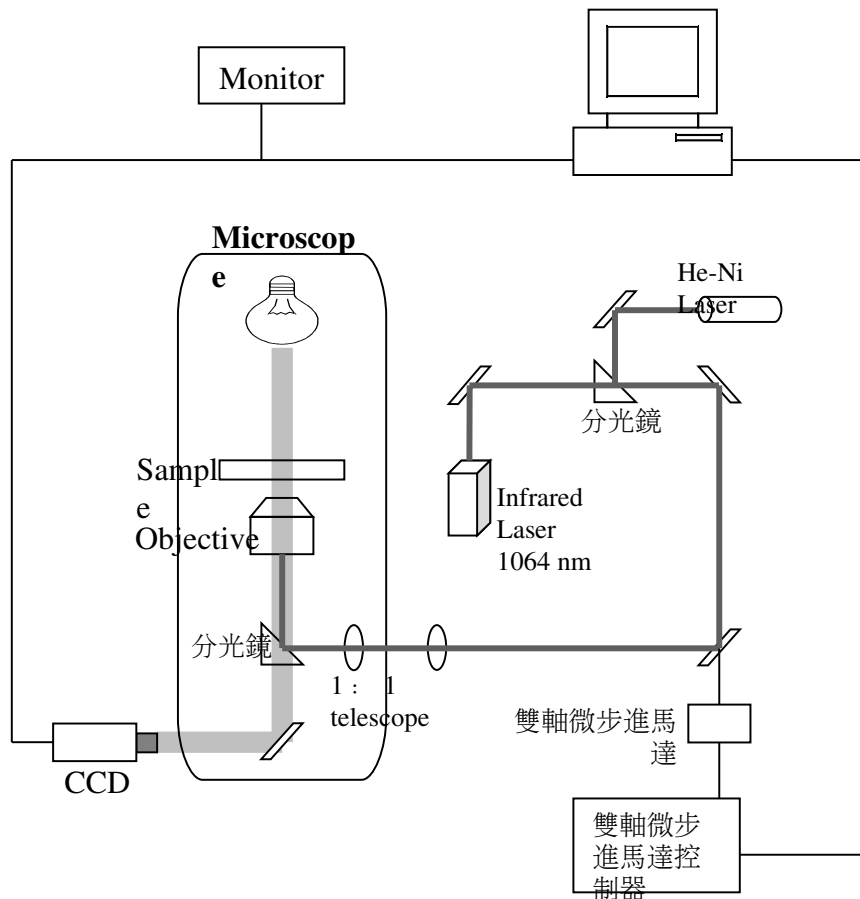
$$k \simeq \frac{1}{N} (N - n_{kill})(N - n_{kill} - 1) P_o,$$

- Well fitted by taking $f \sim a + b k$, with a small.
- $f \sim k$



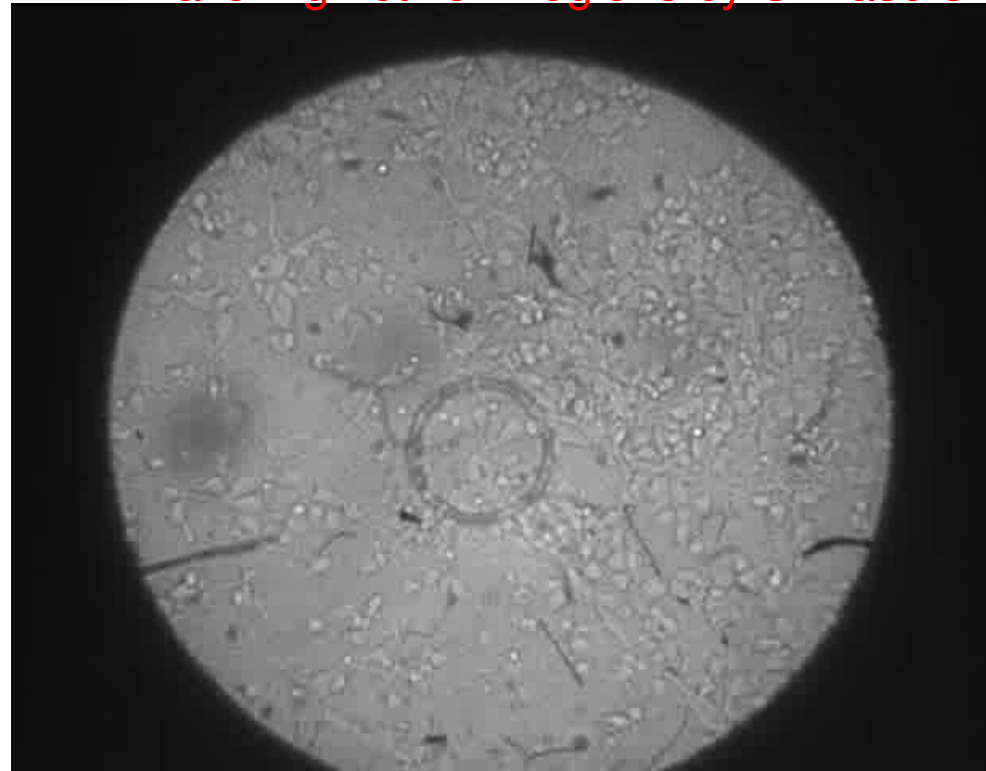
Use synchronized firing freq. to probe the Growth behavior of the network

Manipulating/attacking the neuronal network



Optical Tweezers

Tailoring network regions by UV lasers



Network attack: random or target attack
Network robustness
Regenerative & Re-routing behavior

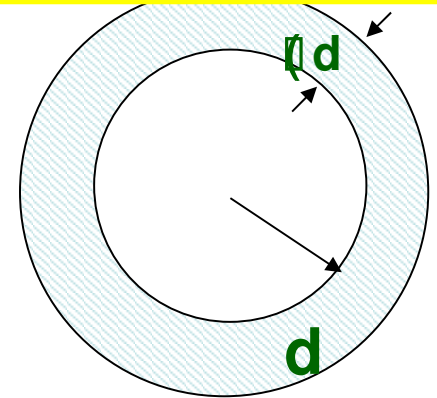
Model for Neuronal Network Growth

Phys.Rev. E 2006

k =mean connectivity in domain of radius d

$P(k)$ = prob. of connecting 2 neurons

ρ =mean cell density



Synchronized Cluster occurs at $t=t_c$ with $f=f_c$

Increase in connectivity: $\Delta k = 2\rho \Delta d \Delta P(k)$

$$d = ut \quad t < t_c$$

Enhanced growth towards synchronized cluster (active search) for $t < t_c$

Experimentally: SF occurs at $t=t_c$ with $f=f_c$

f_c is indep. of ρ : $\square\square$

Assume $f_c \sim kc$, \square SF occurs when sufficient connections are made:

$$kc \sim \rho t_c^2 \quad t_c \propto \frac{1}{\sqrt{\rho}}$$

Retarded growth for $t > t_c$

Empirical result : $(f \sim k) \quad k = k_c + k_o \ln\left(\frac{t}{t_c}\right) \quad t > t_c,$

At $t \sim t_c$ ($k \sim k_c$), the neurons have made enough connections among themselves and cooperativity begins:- a neuron gets enough signals from other neurons so that it surmounts its threshold for further fast growth.

It knows that there are enough connections for cooperativity and there is no need for further increase of connection. Thus the rate of growth R starts to decay.

$$\mathcal{R} \equiv dk/dt = \frac{k_o}{t_c} \exp\left[-\frac{k - k_c}{k_o}\right] \quad \text{for } k \geq k_c.$$

Slowing down for $t > t_c$

$$k = k_c + k_o \ln\left(\frac{t}{t_c}\right) \quad t > t_c,$$

Using diffusive search model: $d^2 = Dt$

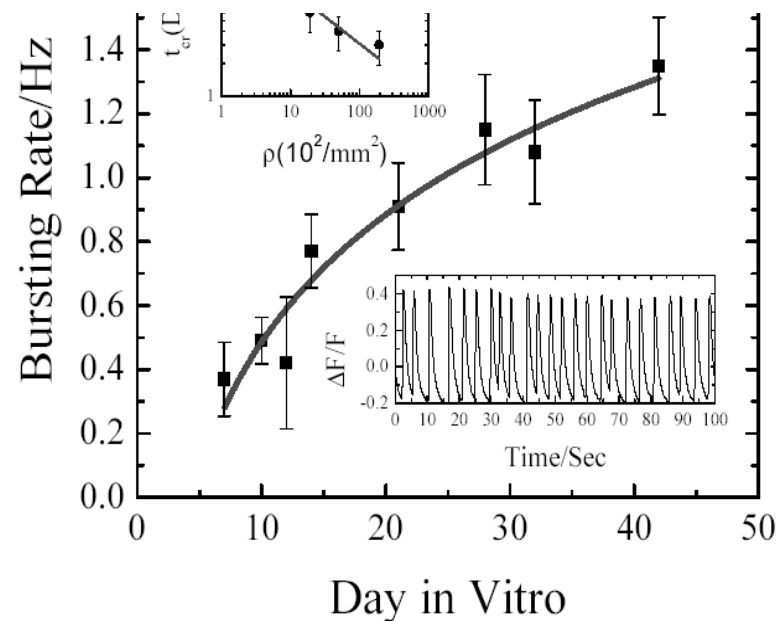
$$\rightarrow P(k) = P_c \exp[-(k - k_c)/k_o] \quad k > k_c$$

$$\delta k = 2 \frac{dk}{k} = 2 \frac{dP(k)}{P(k)}$$

Retarded growth for $t > t_c$:

$$\mathcal{R} \equiv dk/dt = \frac{k_o}{t_c} \exp\left[-\frac{k - k_c}{k_o}\right] \quad \text{for } k \geq k_c.$$

$$\frac{d\mathcal{R}}{dk} = -\frac{\mathcal{R}}{k_o} \quad k > k_c.$$



Expt: $f = f_c + f_0 \log(t/t_c)$

Assume $k/k_c = f/f_c$:

Assume $P(k) \sim 1$ for $k < k_c$

$$\frac{f}{f_c} = \frac{k}{k_c} = \begin{cases} \left(\frac{t}{t_c}\right)^2 & t < t_c \\ 1 + \frac{D}{u^2 t_c} \ln\left(\frac{t}{t_c}\right) & t \geq t_c \end{cases}$$

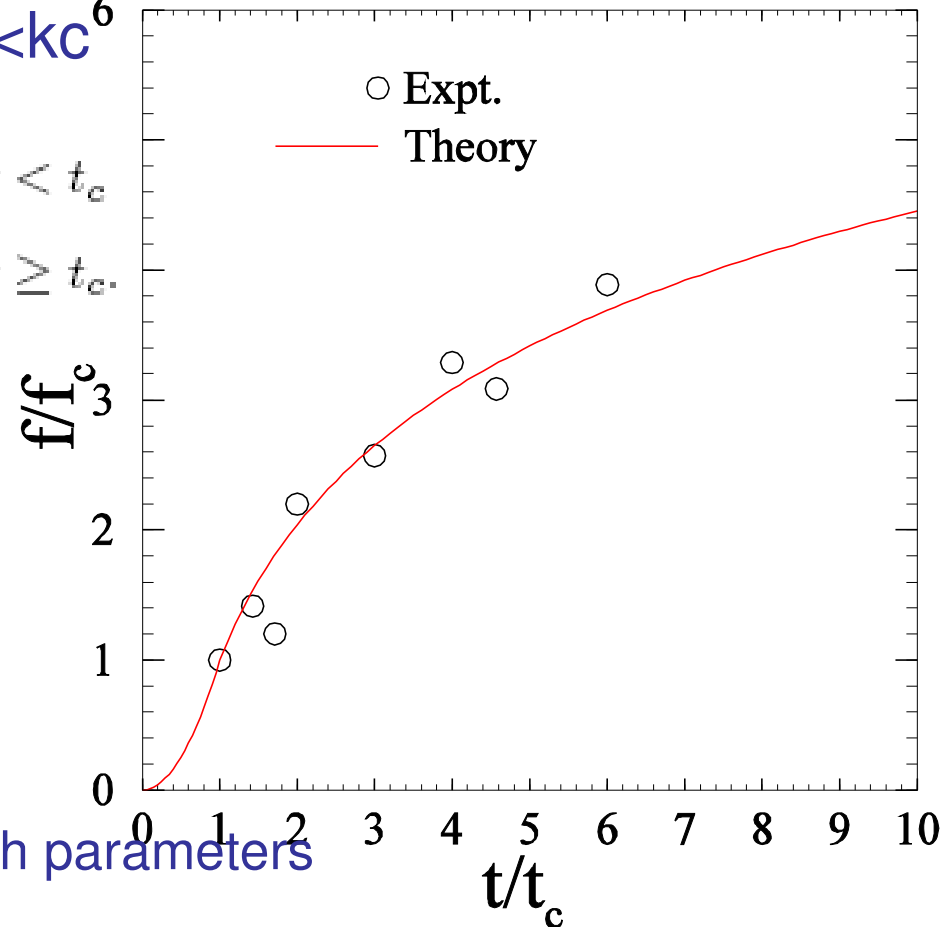
In general with $P(k) = \tilde{P}(k/k_c)$ for $k < k_c$ ⁶

$$\frac{f}{f_c} = \frac{k}{k_c} = \begin{cases} \Phi^{-1}[\Phi(1) \left(\frac{t}{t_c}\right)^2] & t < t_c \\ 1 + \frac{D\Phi(1)P_c}{u^2 t_c} \ln\left(\frac{t}{t_c}\right) & t \geq t_c \end{cases}$$

$$\Phi(x) \equiv \int_0^x \frac{dx}{\tilde{P}(x)}$$

Fitting: $f_0/f_c = k_0/k_c = D/(u^2 t_c) \sim 1.5$

relation between the microscopic growth parameters



Estimates of u and D

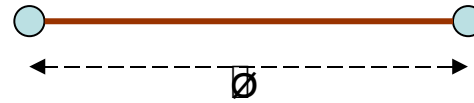
- minimal radius, r_c , of an isolated domain such that SF still occur determined by UV laser tailoring
- For $\rho = 10^4/\text{mm}^2$, we found $r_c \sim 0.15\text{mm}$

$$(ut_c)^2 \rho \sim N_c \sim r_c^2 \rho. \quad D \sim \frac{u^2 t_c f_o}{f_c}$$

- $u \sim 25 \mu\text{m}/\text{day}$; $D \sim 0.0056\text{mm}^2/\text{day}$



Coupling between neurons



- $P(\Delta)$ = mean prob. that 2 neurons initially separated by Δ will be connected

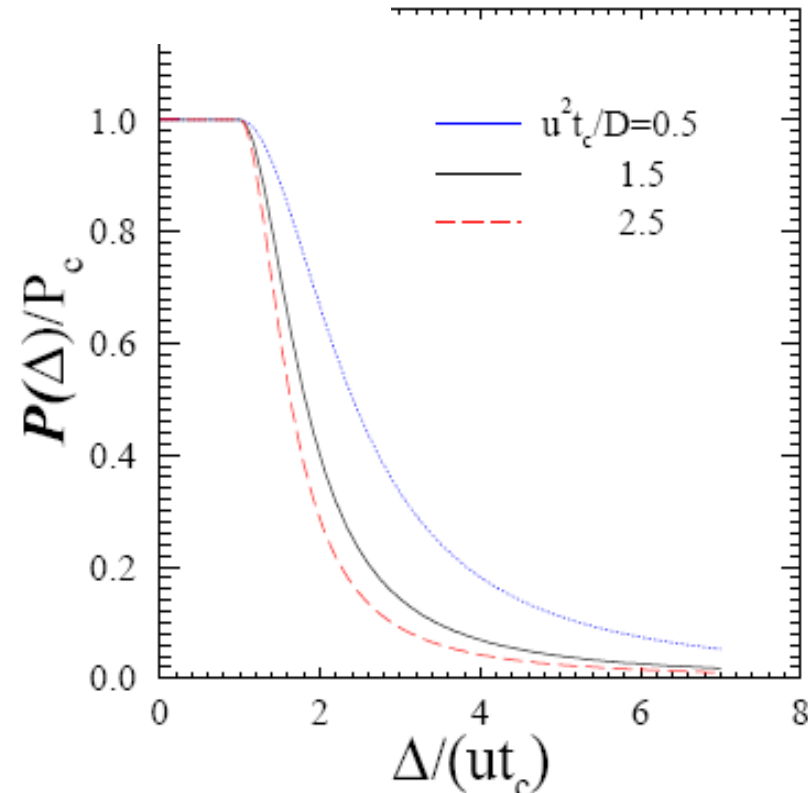
$$\mathcal{P}(\Delta) = \tilde{P} \left(\Phi^{-1} \left(\Phi(1) \left(\frac{\Delta}{ut_c} \right)^2 \right) \right) = \Phi^{-1'} \left(\frac{\pi \rho \Delta^2}{k_c} \right) \quad \text{for } \Delta < ut_c$$

$$\mathcal{P}(\Delta) = \frac{P_c}{1 + \frac{(\Delta - ut_c)^2}{Dt_c}} \quad \text{for } \Delta > ut_c.$$

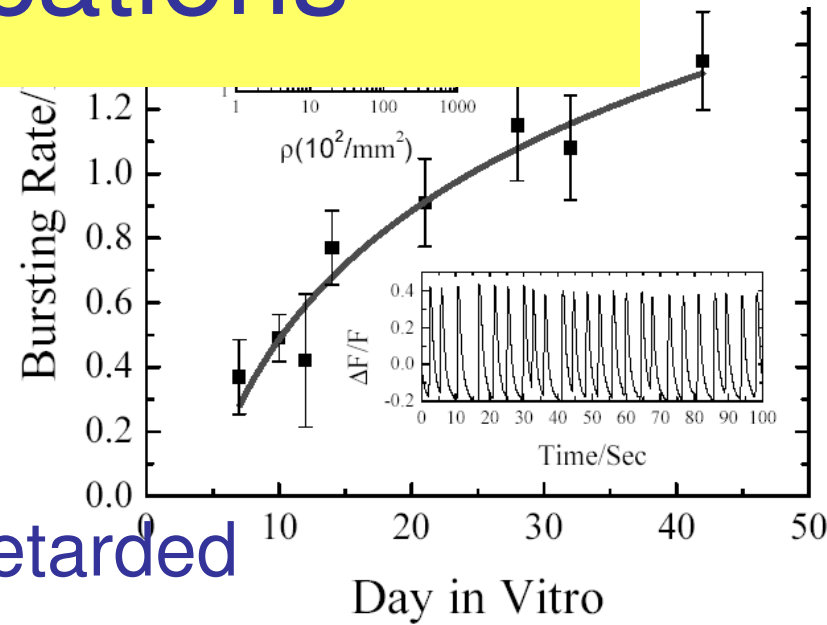
- Characteristic coupling length

$$\Delta_c = ut_c + \sqrt{Dt_c}.$$

- $\Delta_c \sim 0.33\text{mm} \sim 2 r_c$

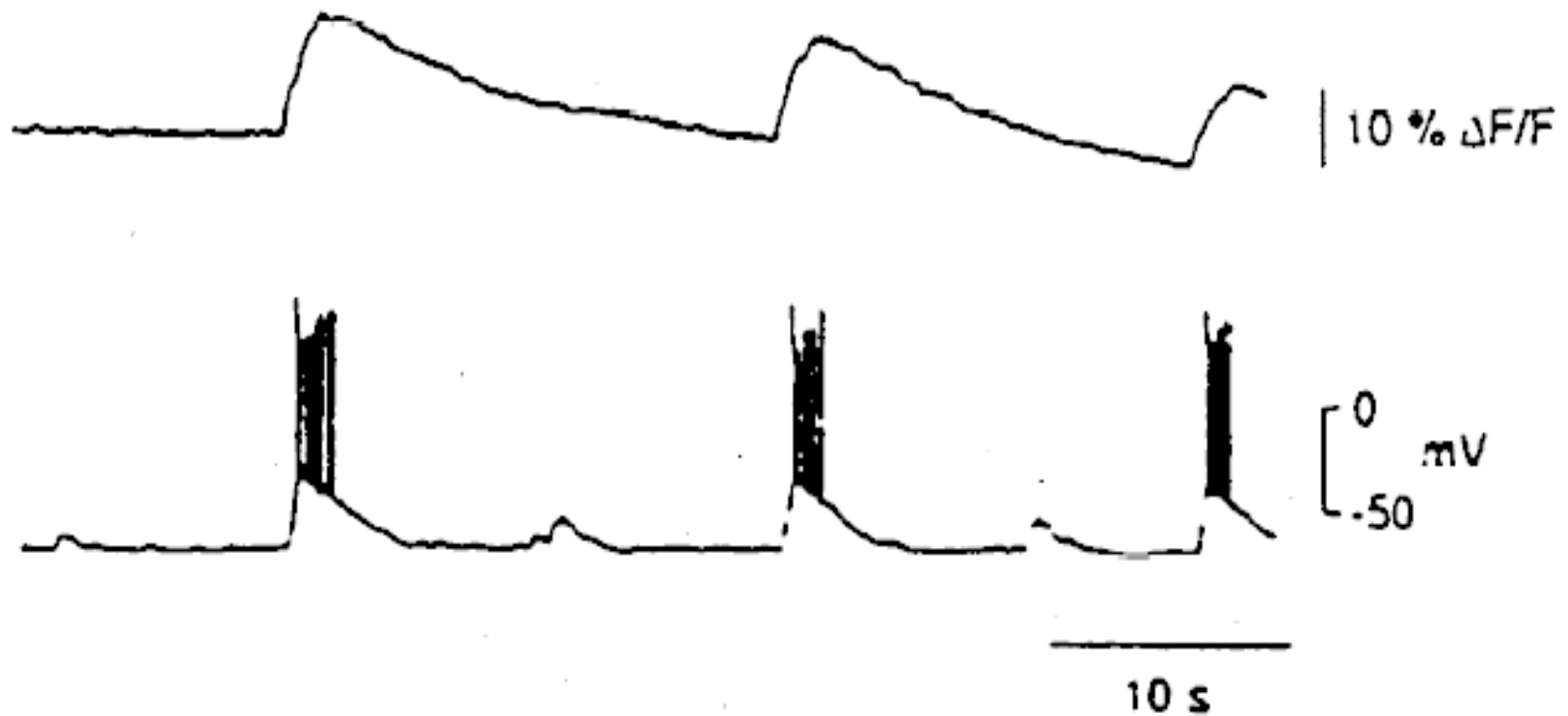


Biological implications

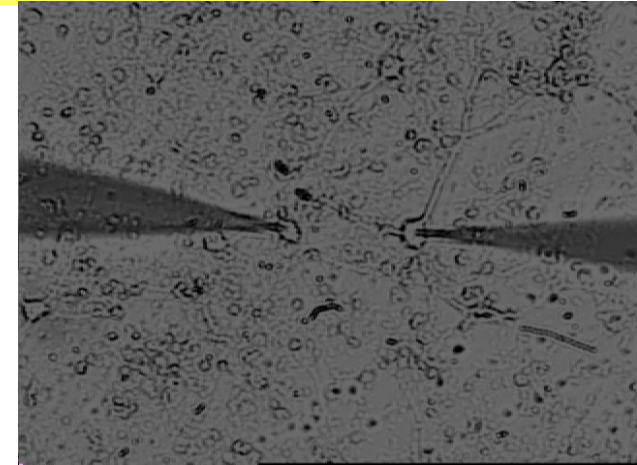


- Active growth in early stage, retarded once goal is achieved.
- Slowing down to maintain a long time span for function: **homeostasis**
- Continuing fast growth used up energy
- Too much connections may exceed information capacity for a single neuron

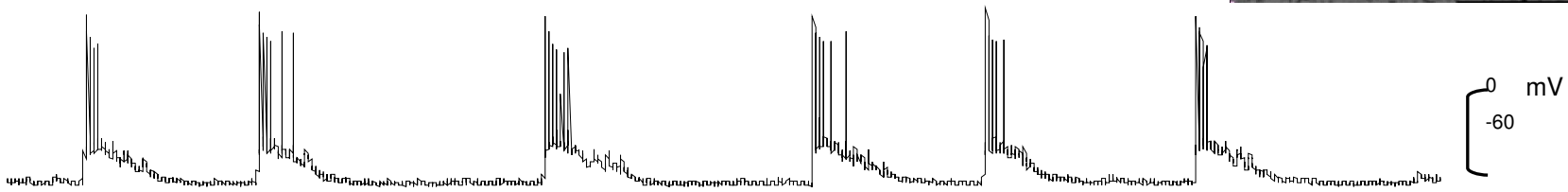
Many Spikes in one pulse: Bursting



Electrophysiology measurement (whole-cell recording, current-clamp)



Glia and neuron mixed culture (8DIV, 5×10^5)



Inter-burst synchronized , but intra-burst is NOT synchronized

2 s

Bursting: role of inhibitory element

- Continuous firing (over excited) is harmful-- excitotoxicity
- Inhibitors (Glia) suppress over-excited neurons
- g =inhibitory field
- Mean-field model
- z =mean connectivity of a neuron to inhibitors

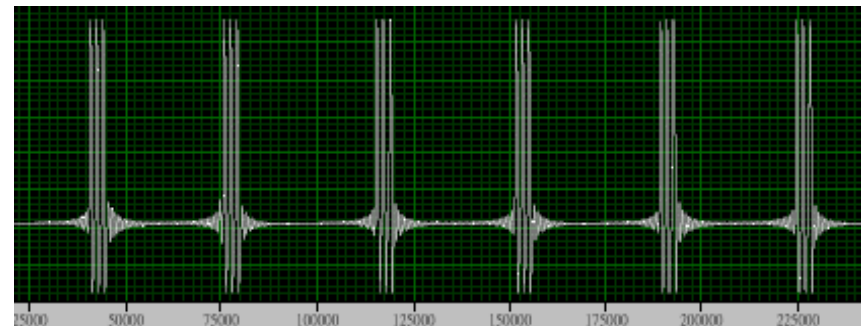
$$\epsilon \frac{dx}{dt} = x - \frac{x^3}{3} - y - zg + \xi(t)$$

$$\frac{dy}{dt} = a + bx - y$$

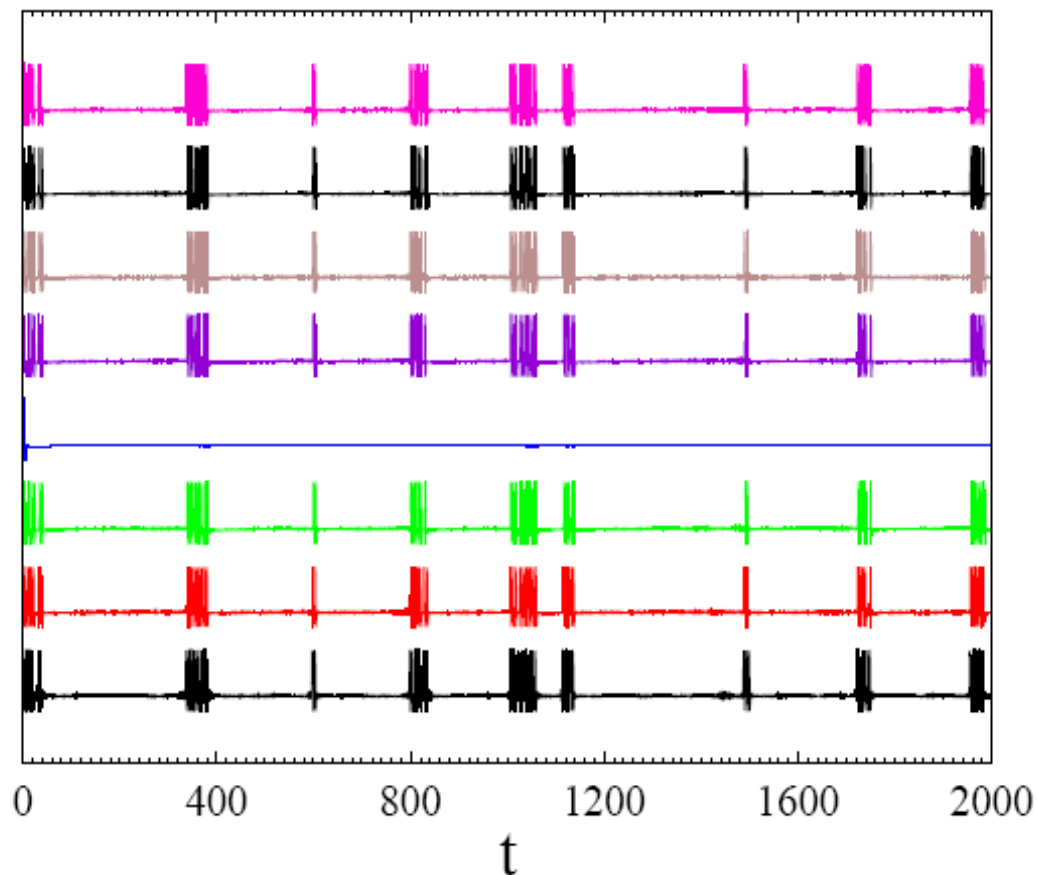
$$\frac{dg}{dt} = -\frac{g}{\tau} + \gamma \Theta(rzx - \theta)$$

$$\Theta(u) = \frac{1}{2} \{1 + \tanh[\alpha u]\}$$

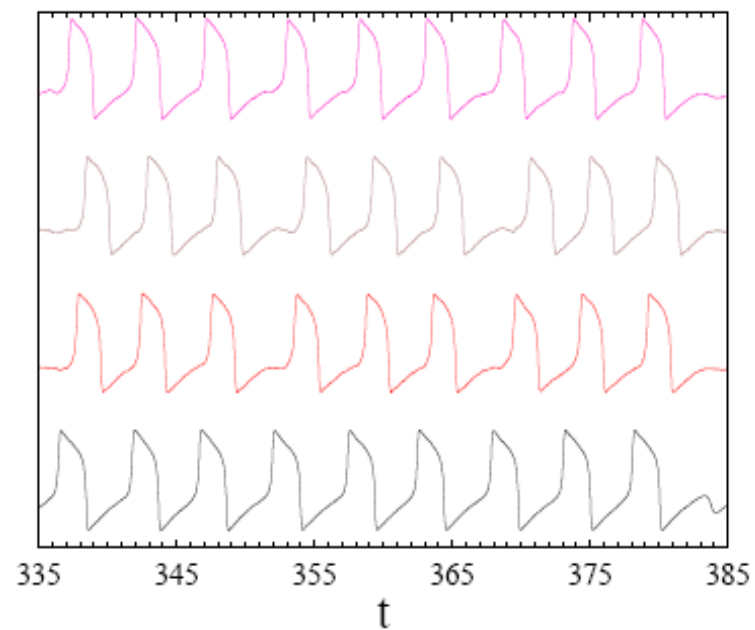
FitzHugh-Nagumo model for neurons



Network of neurons & inhibitors



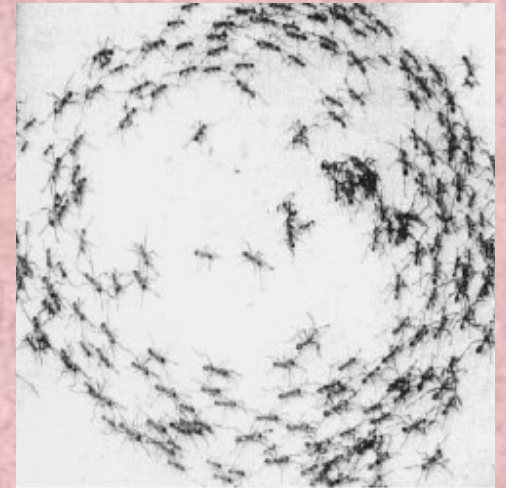
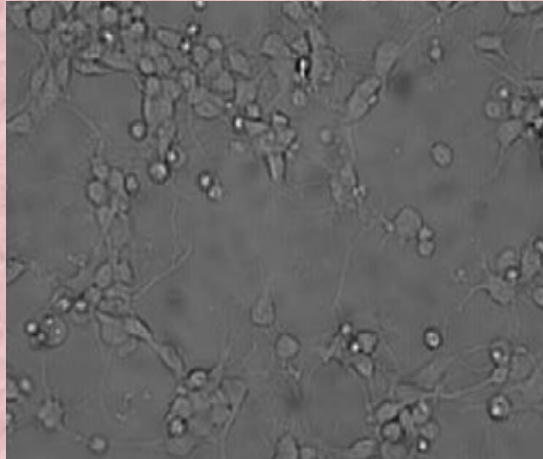
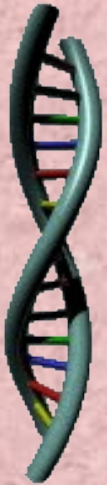
Synchronized Bursting

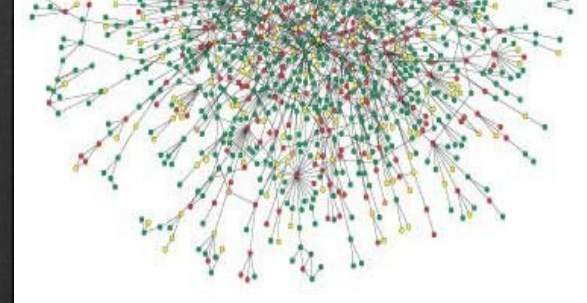
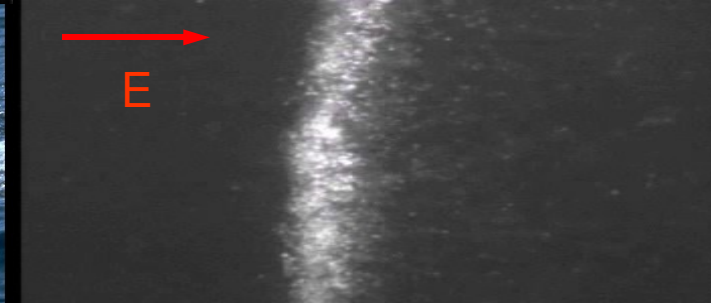
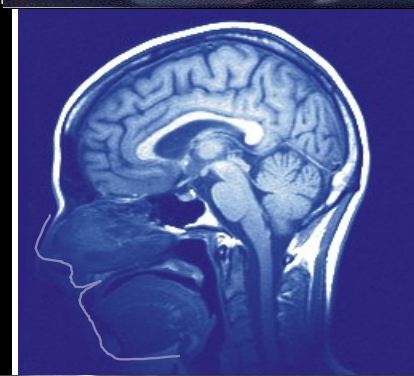
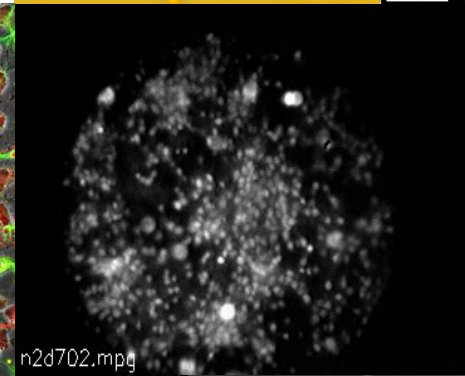
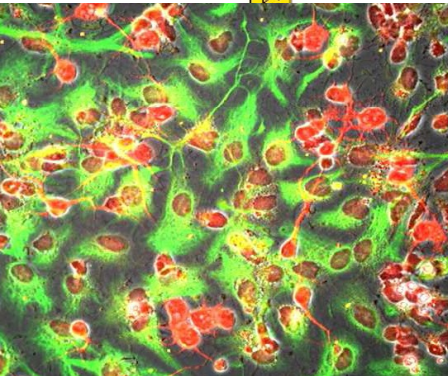
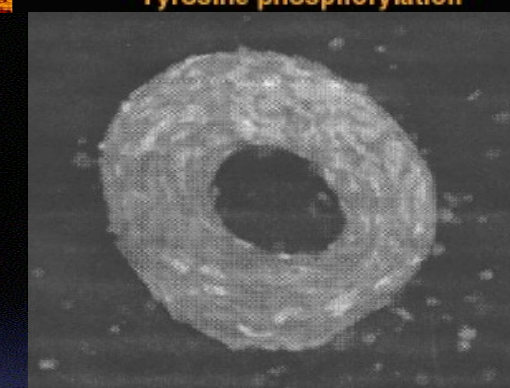
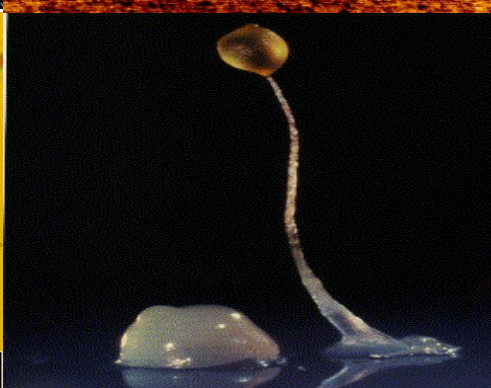
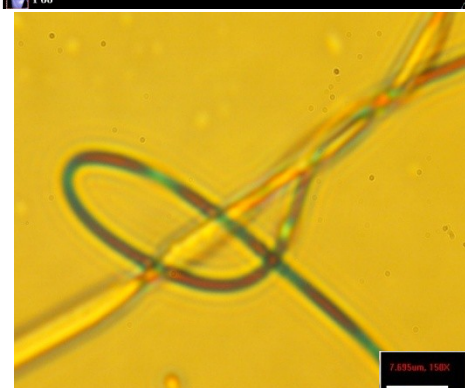
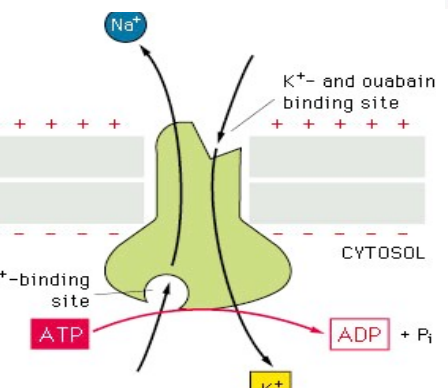
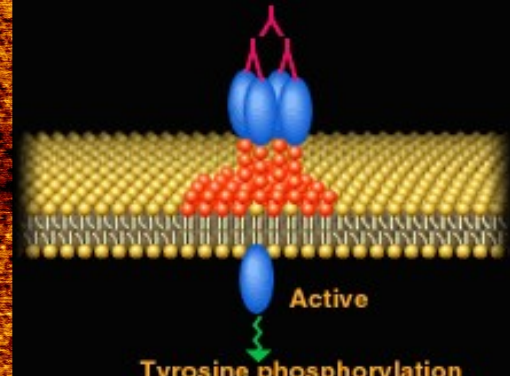
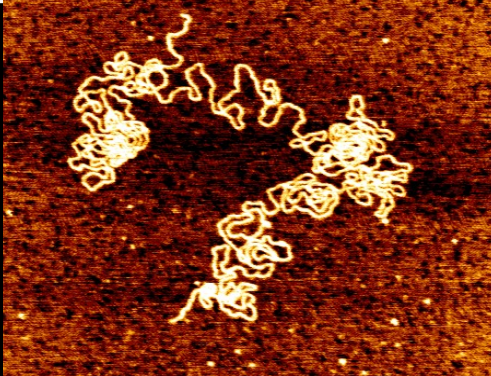
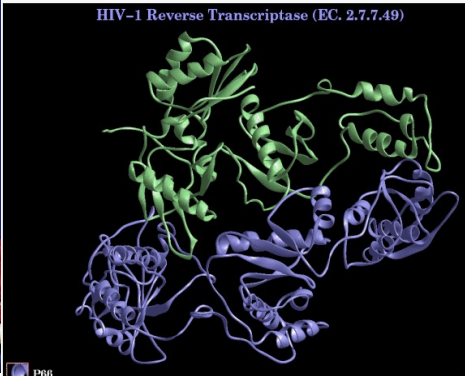


Intra-burst NOT synchronized

- **What Physicists can do in Biology ?**

a lot of interesting and unexplored science
from molecules to collective behavior of organisms





Acknowledgements

Collaborators

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The End